

CLINICAL METAGENOMICS OF HOSPITAL-ACQUIRED PNEUMONIA

S. Hauser, V. Lazarevic, M. Tournoud, E. Ruppé, E. Santiago Allexant, G. Guigon, S. Schicklin, V. Lanet, M. Girard, P. François, C. Mirande, S. Chatellier, G. Gervasi and J. Schrenzel

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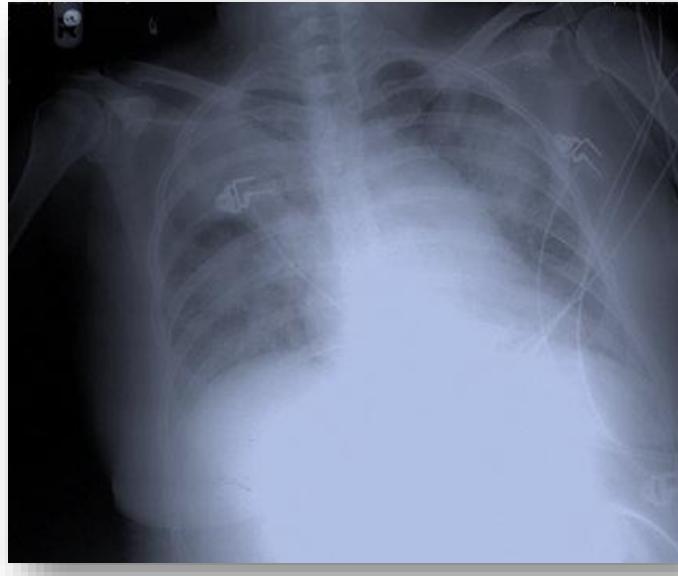
PIONEERING DIAGNOSTICS



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Introduction



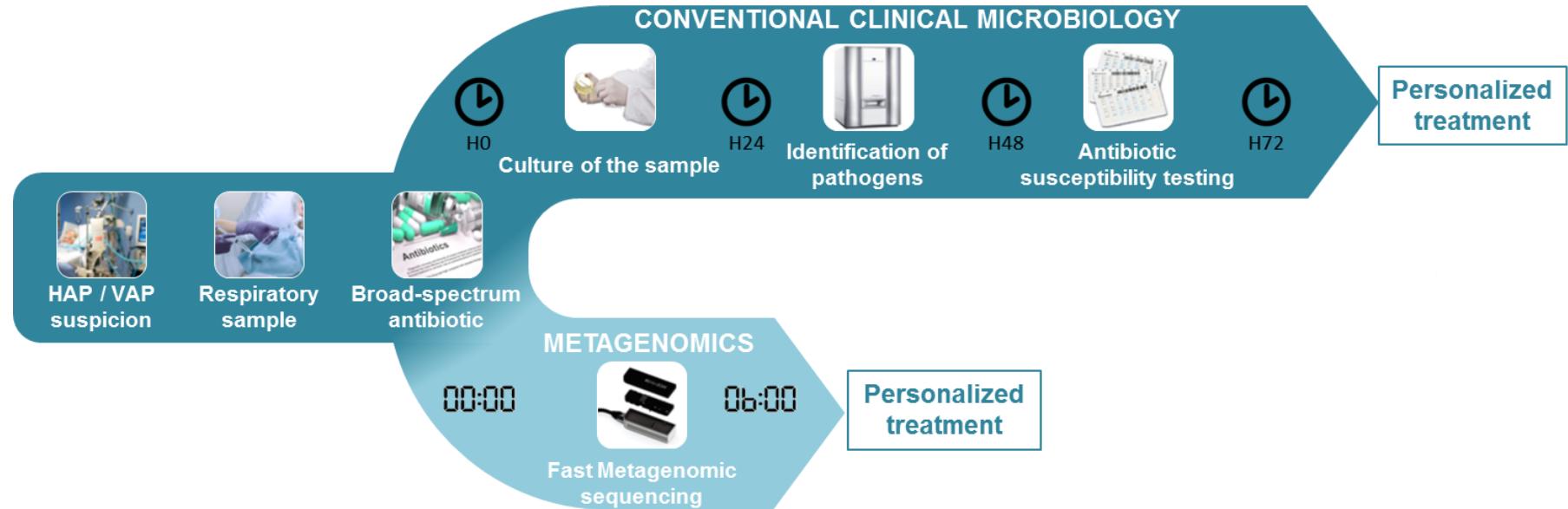
► Medical need to adjust antibiotic treatment

- Rapid identification of pathogens detected above clinical threshold
- Rapid detection of drug resistances

- Hospital-Acquired Pneumonia (HAP) and Ventilator-Associated Pneumonia (VAP)
 - Common nosocomial infections in intensive care units
 - Highest morbidity and mortality rate of all nosocomial infections

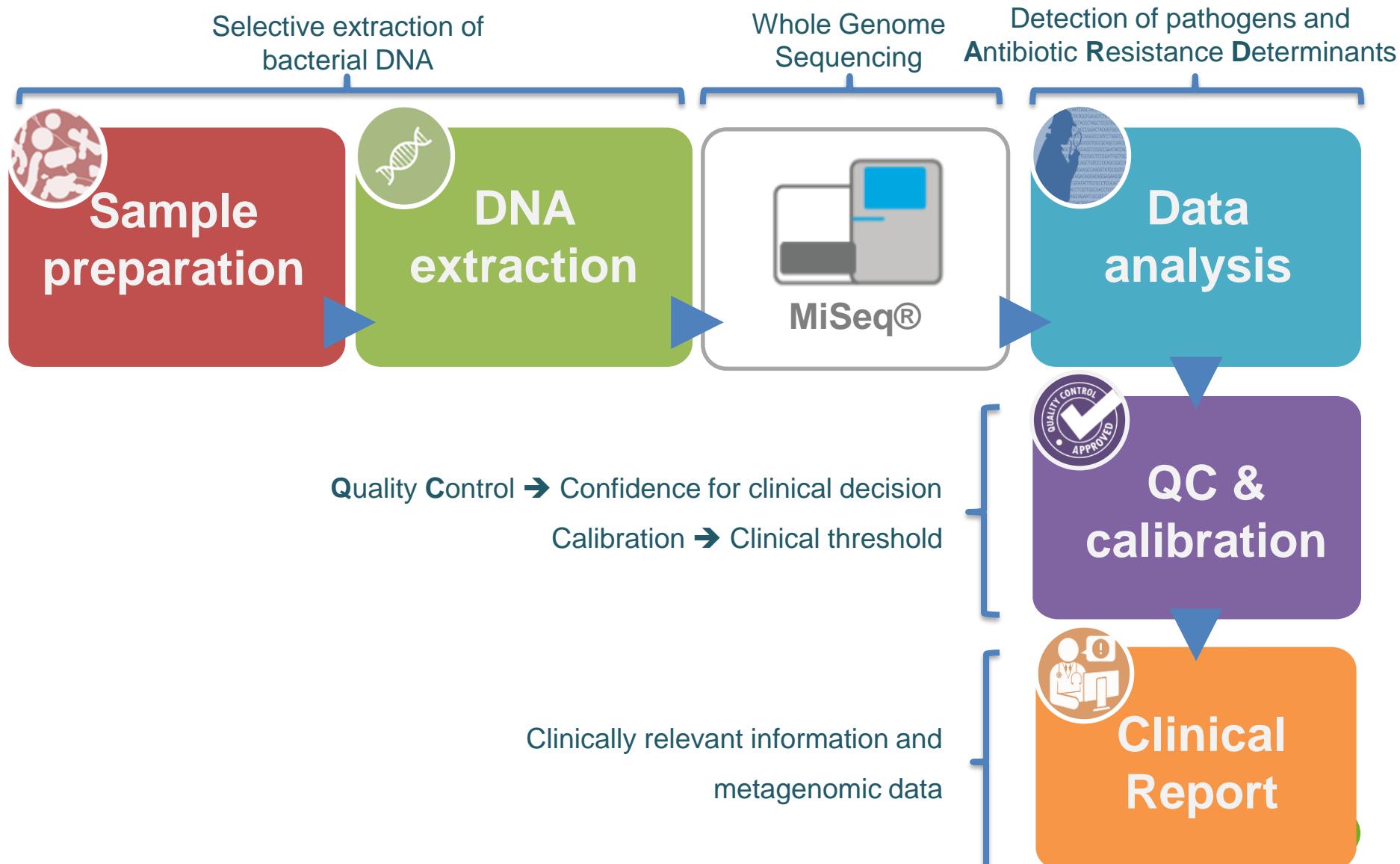


Project Objectives



- ▶ **Bypass the time-consuming culture-dependent methods for identification of pathogens and resistance profiles in 6 hours**
- ▶ **Whole Genome Sequencing for exhaustive detection of bacteria :**
 - ▶ Conversion of metagenomics data into clinically actionable information
 - ▶ Quantification of pathogens to differentiate bacterial colonization from infection
 - ▶ Detection of Antibiotic Resistance Determinants and association to detected pathogens

Clinical Metagenomics Workflow for BronchoAlveolar Lavage (BAL) and mini-BAL



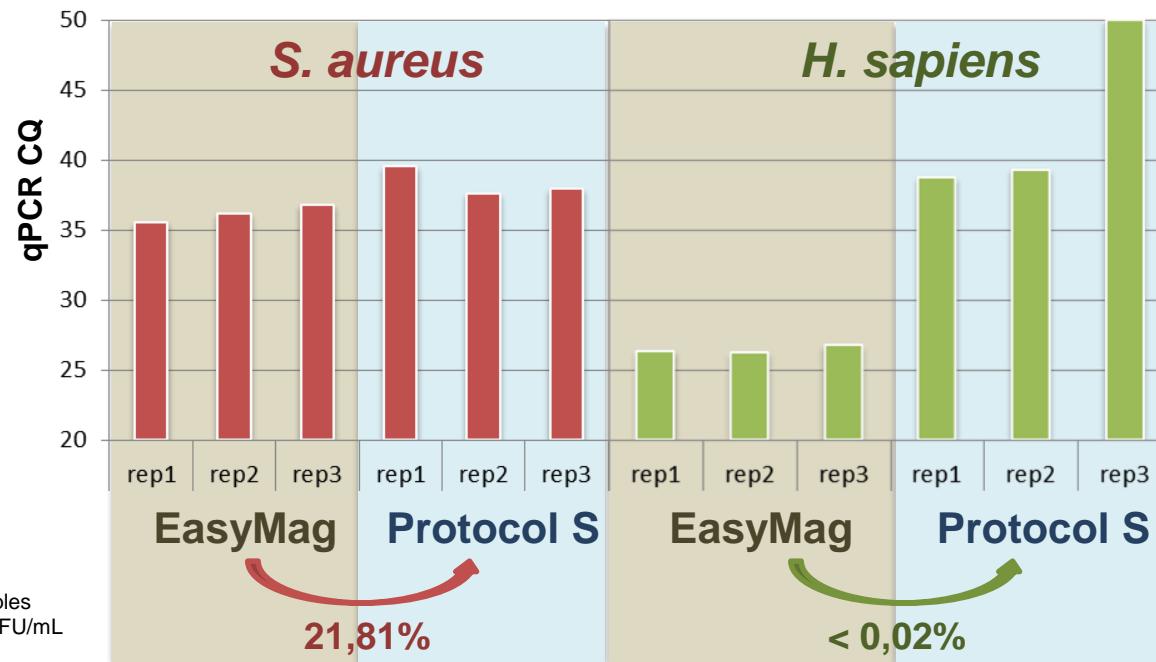
Improved sensitivity by specific sample preparation

► Clinical samples are BAL or mini-BAL

- Contain huge quantity of DNA from patient → Impact detection sensitivity and costs
- Bacterial concentrations ranging from $<1^{E}2$ CFU/mL to $>1^{E}6$ CFU/mL

► Selective sample preparation

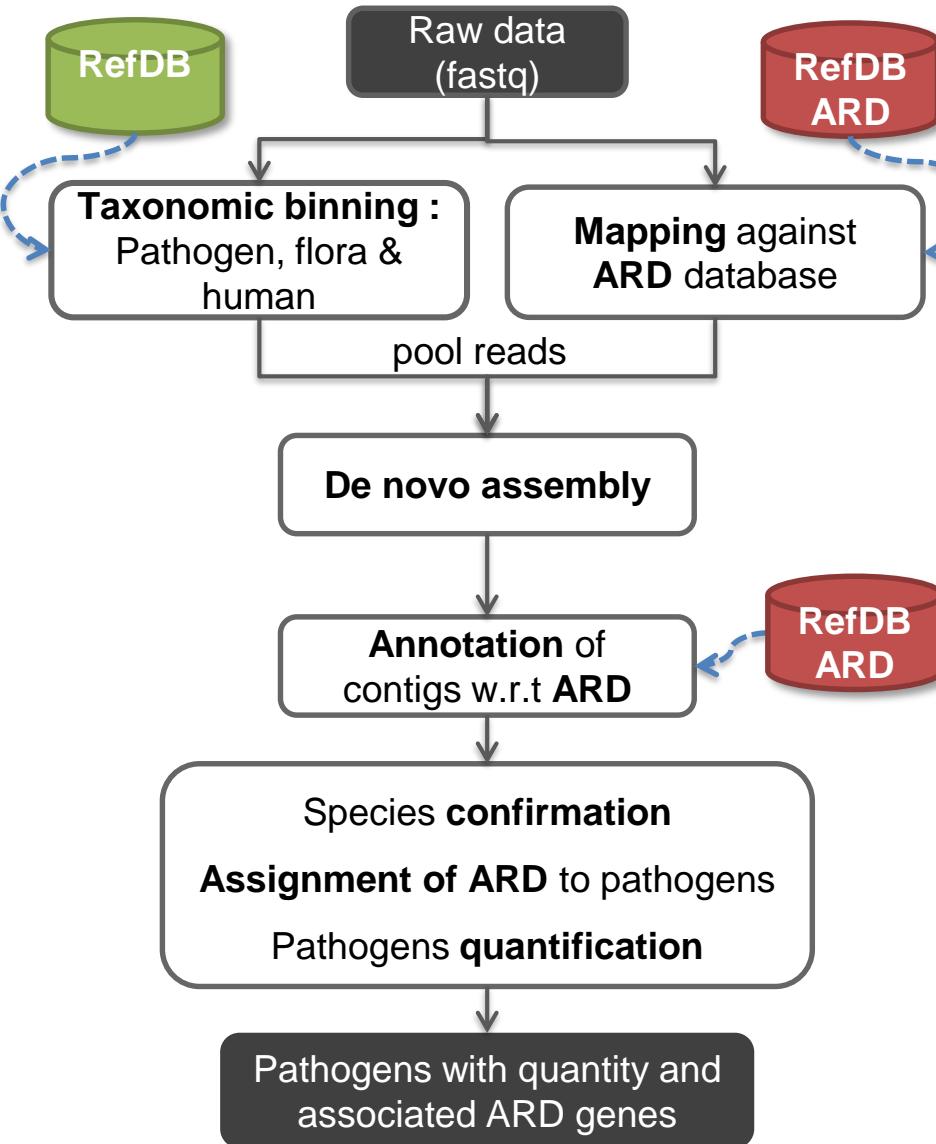
- Protocol S eliminates 99,98 % of human DNA in BAL samples



Increase in
bacterial/Human
DNA ratio
 $> 1\,000 X$

→ Improvement
of pathogen
detections

Dedicated bioinformatics pipeline to extract clinical information



- ▶ **Dedicated DataBase construction**
- ▶ **Separation of taxonomic binning and Antibiotic Resistance Determinants mapping**
- ▶ **Extraction of clinical information:**
 - ▶ Pathogens identification with specificity checking (16S, MetaPhlAn, BLAST)
 - ▶ Quantification of pathogens
 - ▶ Association of ARD to pathogens
 - ▶ Possible → Resistant pathogens
 - ▶ Not possible → Resistant flora ?
 - ▶ ARD not present → Sensitive pathogens

Quality control to facilitate clinical decision with confidence



Draft - Not for Implementation

1 **Infectious Disease Next Generation
2 Sequencing Based Diagnostic Devices:
3 Microbial Identification and Detection
4 of Antimicrobial Resistance and
5 Virulence Markers**

6
7 **Draft Guidance for Industry and
8 Food and Drug Administration Staff**

9
10 **DRAFT GUIDANCE**

11
12 This draft guidance document is being distributed for comment purposes
13 only.

14
15 Document issued on: May 13, 2016

► **Sample Processing Control
(SPC) designed according to
FDA guidance**

- Spiked at sufficient concentrations into each clinical sample
- Analyzed simultaneously with the clinical targets

► **FastQC**

- Sequence reads quality control

► **Specificity checking**

- Control correct sequence binning of pathogenic bacteria

Correlation of pathogen detection between culture and metagenomics

► Culture and sequencing detections match at 96,4%

► Less than 0,5% of “false negative”

- ▶ Observed in multi-infected patient → competition effect
 - ▶ No sample would be declared as negative !

► 3,1 % of “false positive”

- ▶ Better sensitivity of NGS
 - ▶ In sample 7, presence of *S. aureus* DNA was confirmed by qPCR

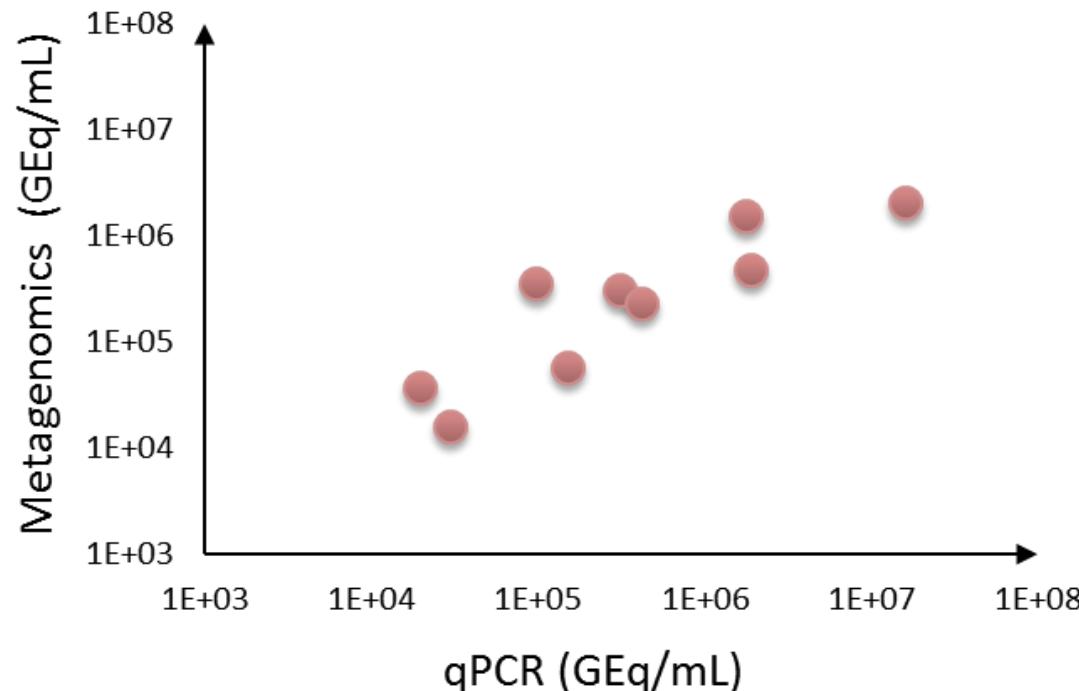
Metagenomics		
Culture	+	-
+	25	5
-	29	891

Calibration for quantification of bacteria around clinical threshold

► Quantitative detection

- Internal calibration and software allow for quantifying bacterial genomes in **Genome Equivalents (GEq) unit.**
- Metagenomics quantification correlates with real-time PCR

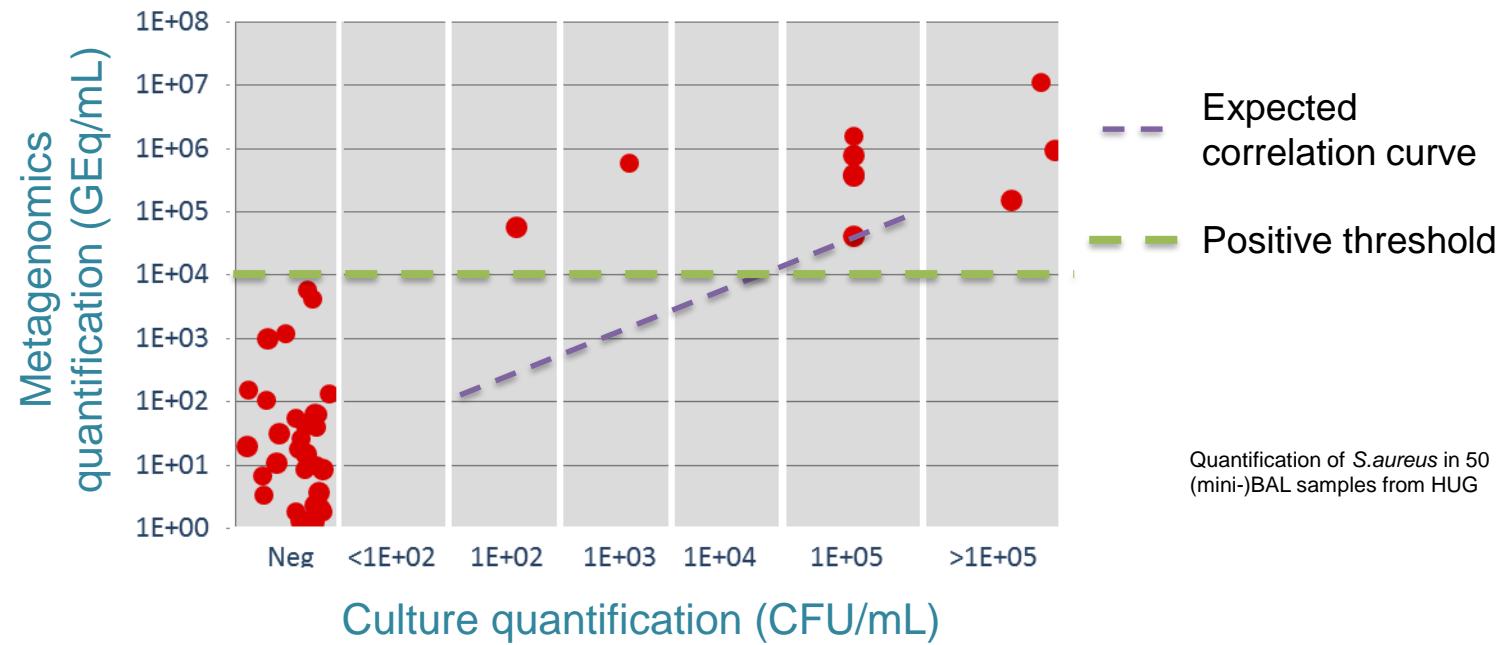
**Quantification of
S. aureus in BAL
samples by shotgun
sequencing and real-
time PCR (qPCR)**



Calibration for quantification of bacteria around clinical threshold

► Bacterial colonization or infection ?

- Clinical threshold at $1\text{E}4$ CFU/mL for BAL samples and $1\text{E}3$ CFU/mL for mini-BAL
- No direct correlation between CFU and quantified genomes in clinical samples
- However, culture positive and negative samples can be distinguished by genome quantification



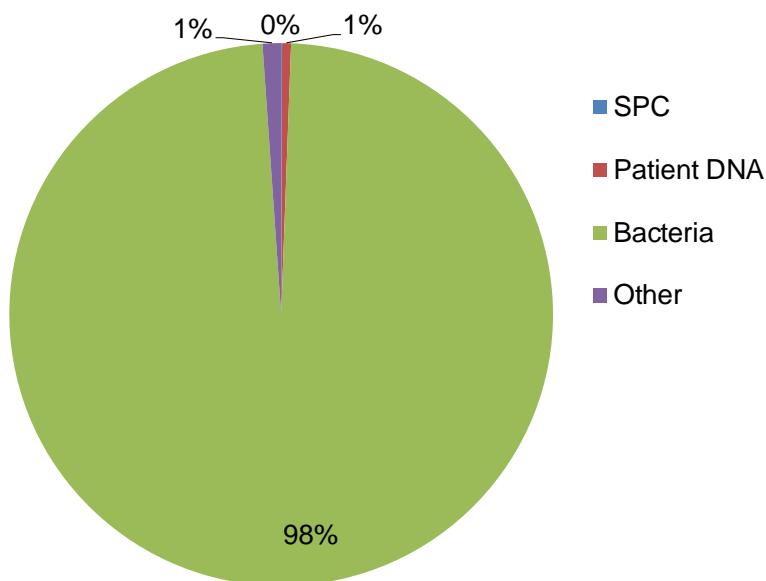
→ Needs to adjust clinical threshold when expressed in GEq/mL 10

Clinical reporting of metagenomics data

BAL_107

Sample Processing Control
SPC not detected, however <i>C. freundii</i> and <i>K. pneumoniae</i> are detected above the clinical threshold

DNA content



Species Of Interest	% Reads tot.	Concentration	Specificity control
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<i>Acinetobacter baumannii</i>	0,01%		
<i>Citrobacter freundii</i>	64,15%	7,1E+06 GEq/mL	YES
<i>Escherichia coli</i>	1,17%		NO
<i>Haemophilus influenzae</i>	0,00%		
<i>Klebsiella oxytoca</i>	0,37%		NO
<i>Klebsiella pneumoniae</i>	14,63%	1,6E+06 GEq/mL	YES
<i>Pseudomonas aeruginosa</i>	0,00%		
<i>Serratia marcescens</i>	0,01%		
<i>Staphylococcus aureus</i>	0,00%		
<i>Stenotrophomonas maltophila</i>	0,00%		
<i>Streptococcus pneumoniae</i>	0,00%		

Antibiotic Resistance Determinants

	CMY_116	LEN-9
<i>Citrobacter freundii</i>	+	
<i>Klebsiella pneumoniae</i>		+

Positive detection

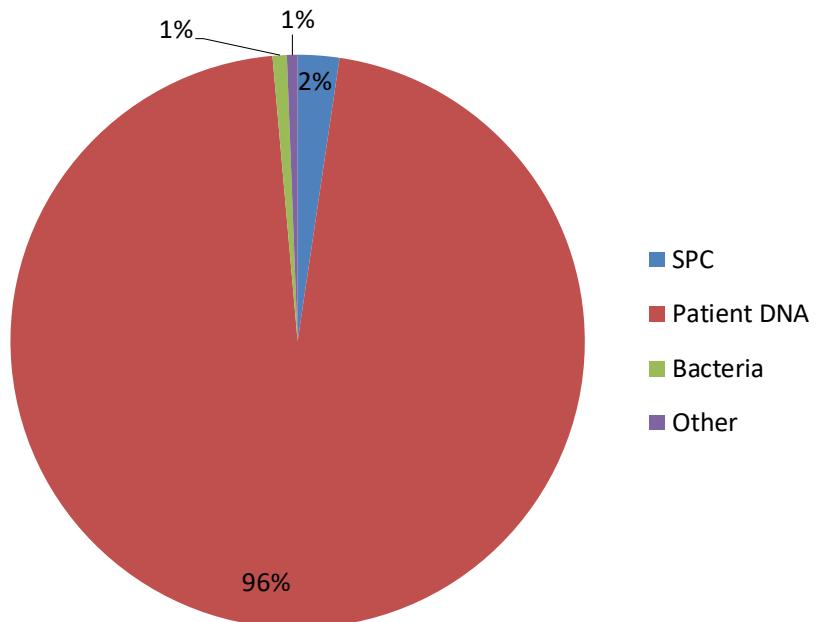
Clinical reporting of metagenomics data

mini-BAL_6

Sample Processing Control

SPC is detected, the limit of detection (7,1E+02 GEq/mL) is below the clinical threshold.

DNA content



Species Of Interest

Species	% Reads tot.	Concentration	Specificity control
<i>Acinetobacter baumannii</i>	0,00%	NEG	
<i>Citrobacter freundii</i>	0,00%	NEG	
<i>Escherichia coli</i>	0,00%	NEG	
<i>Haemophilus influenzae</i>	0,00%	NEG	
<i>Klebsiella oxytoca</i>	0,00%	NEG	
<i>Klebsiella pneumoniae</i>	0,00%	NEG	
<i>Pseudomonas aeruginosa</i>	0,00%	NEG	
<i>Serratia marcescens</i>	0,00%	NEG	
<i>Staphylococcus aureus</i>	0,00%	NEG	
<i>Stenotrophomonas maltophili</i>	0,00%	NEG	
<i>Streptococcus pneumoniae</i>	0,00%	NEG	

Antibiotic Resistance Determinants

No detection

Negative detection

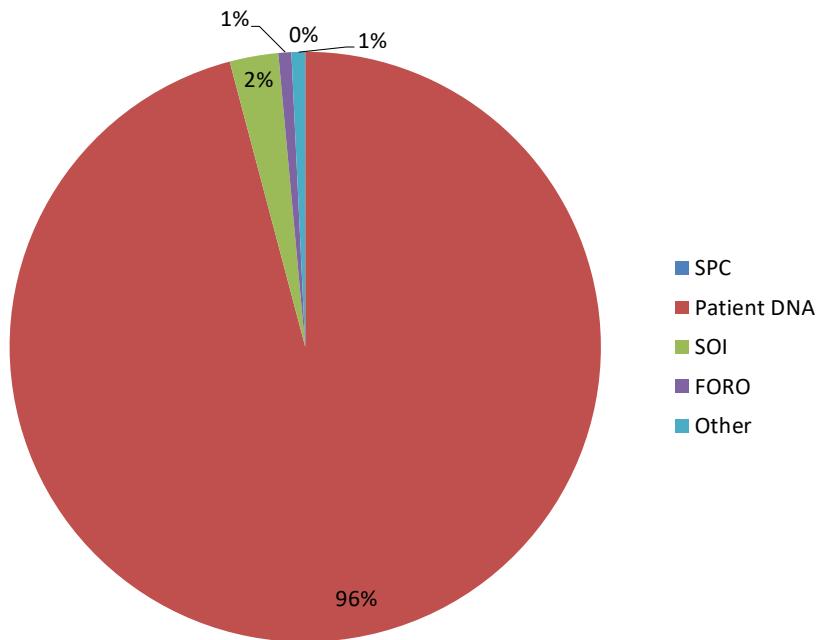
Clinical reporting of metagenomics data

mini-BAL_253

Sample Processing Control

SPC was not detected, the process did not meet the expected efficiency for safe interpretation of results

Microbial Documentation



Species of interest			
Species	% Reads tot.	Concentration	Confirmation
<i>Acinetobacter baumannii</i>	0,00%	?	
<i>Citrobacter freundii</i>	0,00%	?	
<i>Escherichia coli</i>	0,02%	?	
<i>Haemophilus influenzae</i>	0,00%	?	
<i>Klebsiella oxytoca</i>	0,00%	?	
<i>Klebsiella pneumoniae</i>	0,02%	?	
<i>Pseudomonas aeruginosa</i>	0,00%	?	
<i>Serratia marcescens</i>	0,00%	?	
<i>Staphylococcus aureus</i>	0,00%	?	
<i>Stenotrophomonas maltophila</i>	0,00%	?	
<i>Streptococcus pneumoniae</i>	0,00%	?	

Antibiotic Resistance Determinants

?

Invalid detection

Conclusion and perspectives

Pathogens detection



- ▶ Validation of the metagenomics workflow
- ▶ Inclusion of new centers

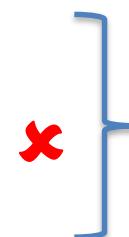


Pathogens quantification

Sample Processing Control

ARD detection

ARD association to pathogens



- ▶ Automation of sample preparation
- ▶ Continuous evolution of NGS platform :
 - ▶ Gain in speed
 - ▶ Sequencing depth
 - ▶ Cost reduction

Time To Result



Thank you



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Gaspard GERVASI



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Emmanuelle SANTIAGO ALLEXANT



Stéphane SCHICKLIN



Ghislaine GUIGON



Veronique LANET



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Caroline MIRANDE



Margaux CHAPEL



Sonia CHATELIER



Frédéric LEMAUFF

Asmaà FRITAH-LAFONT

Karen LOUIS

Philippe LEISSNER

Christelle BOISSE

Adrien SALIOU



Jacques SCHRENZEL



Vladimir LAZAREVIC



Etienne RUPPE



Myriam GIRARD



Patrice FRANCOIS



Albrice LEVRAT