

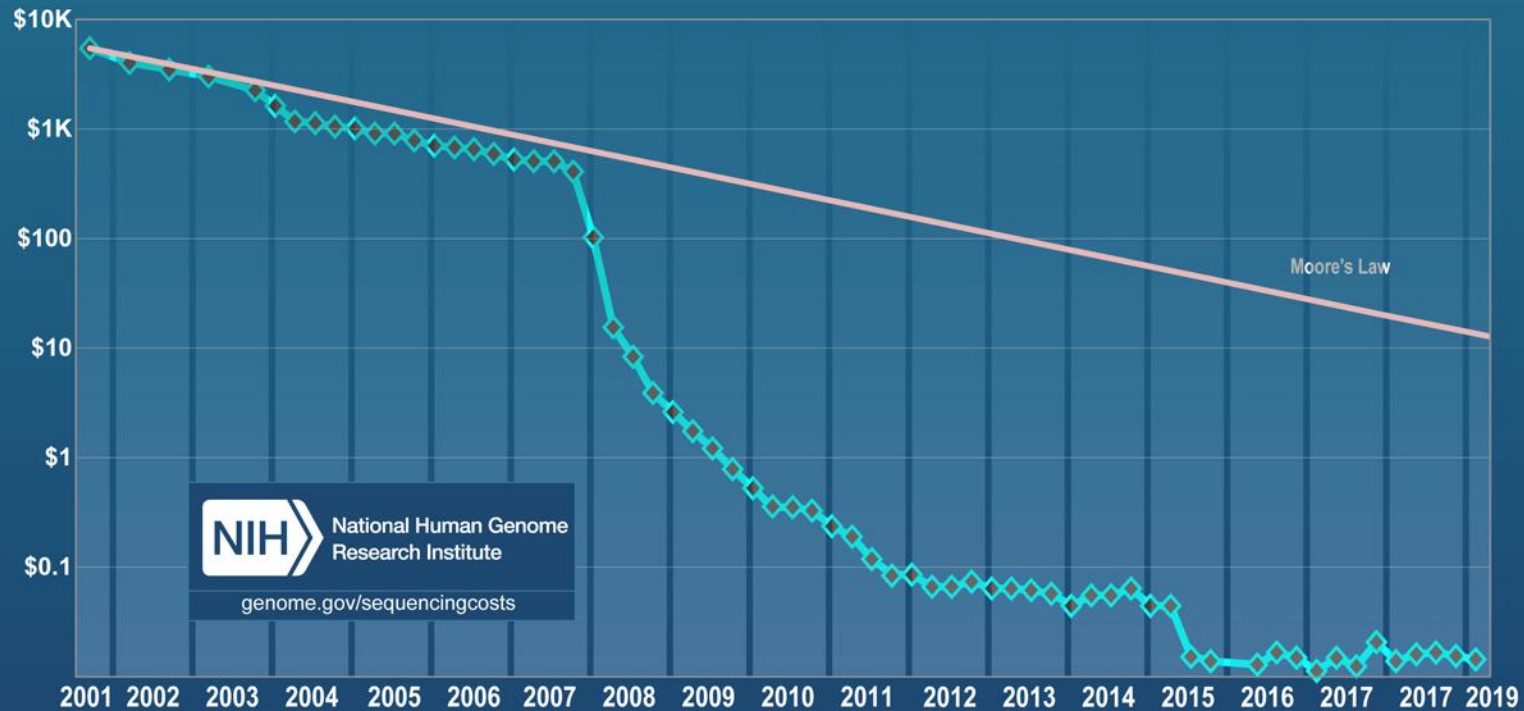
# USING PROXIMITY TO FIX ASSEMBLY

Ivan Liachko, Ph.D.  
Phase Genomics, Inc.  
Seattle, WA, USA

Web: [phasegenomics.com](http://phasegenomics.com)

Twitter: @PhaseGenomics

## Cost per Raw Megabase of DNA Sequence





# SEQUENCE



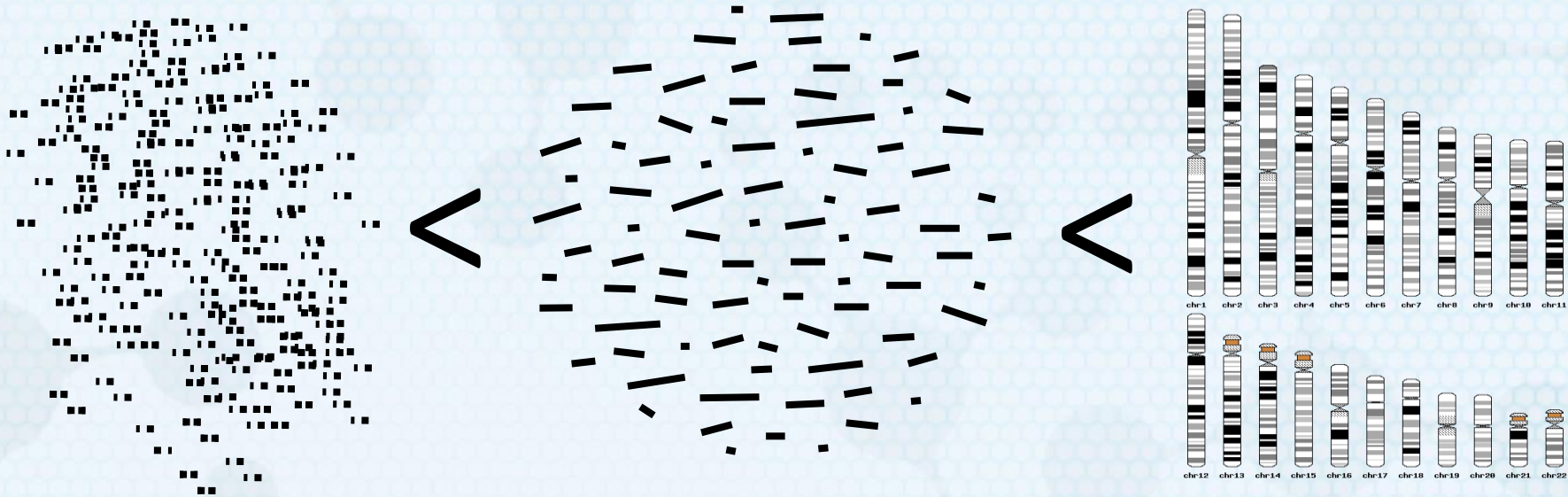
# ALL THE THINGS

memegenerator.net

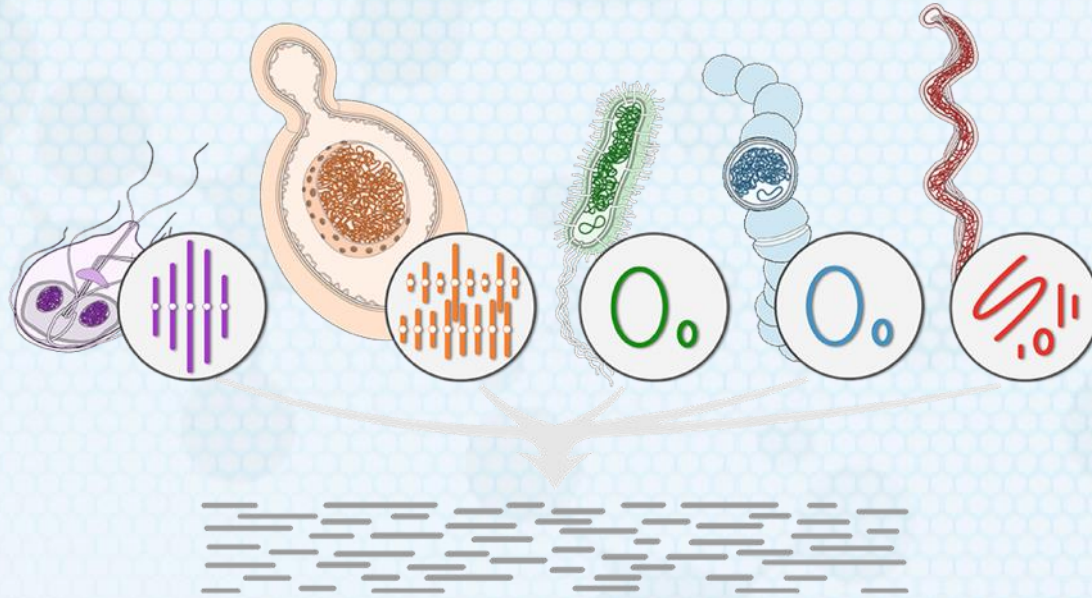




# Reads < Contigs < Genome



# The problem with shotgun metagenomics



- Cannot tell which sequences belong to which organism
- Binning methods are inaccurate, hard to reproduce
- No way to track plasmids / viruses / antibiotic resistance
- Missing lots of organisms that are not in databases

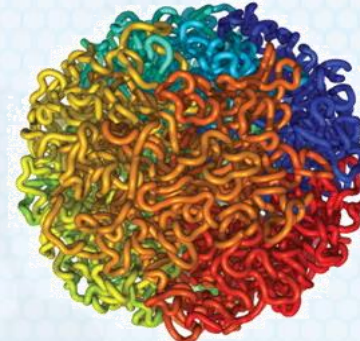
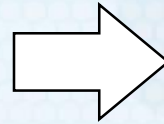
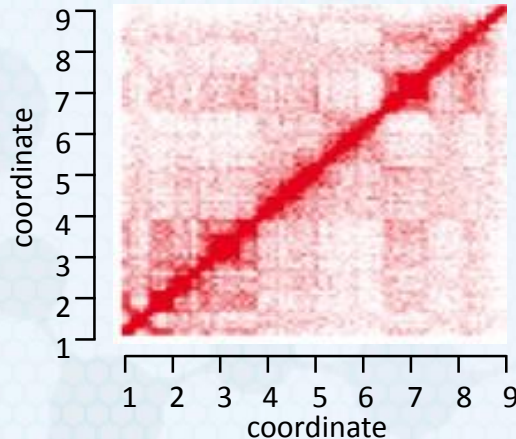
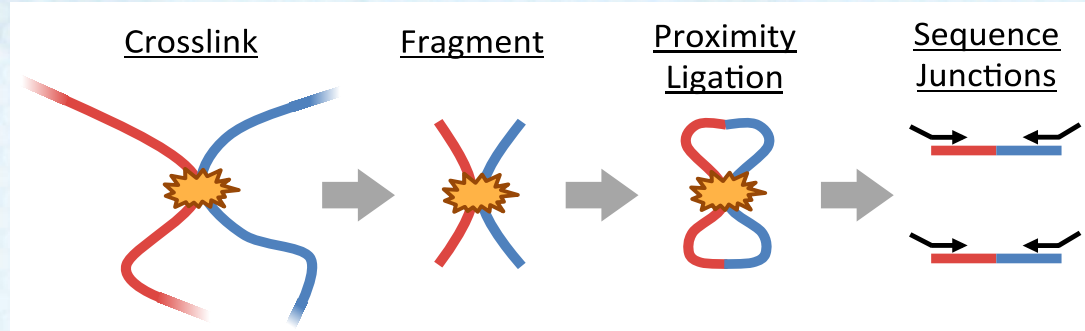


# Genomes are packaged into 3D structures



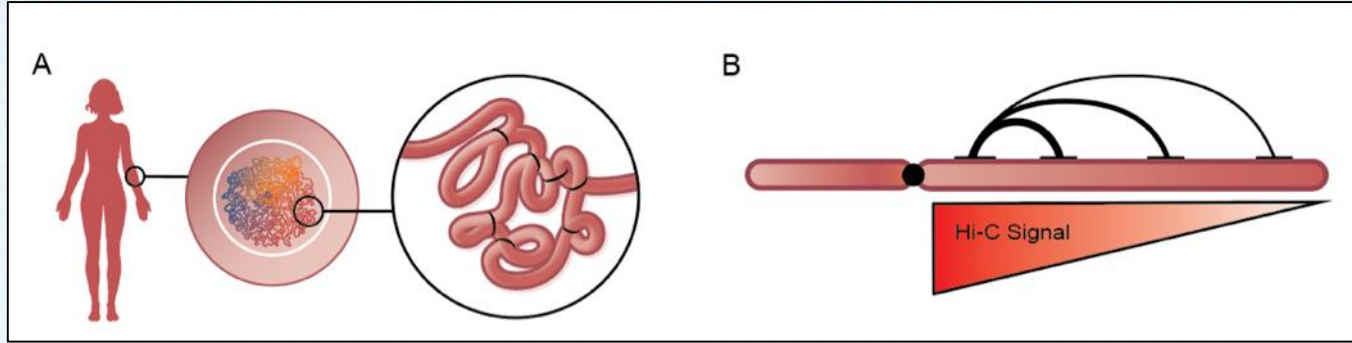
Lieberman-Aiden, *et. al.* Science, 2009

# Proximity Ligation (Hi-C) captures the 3D structure of chromosomes



Lieberman-Aiden, *et. al.* Science, 2009

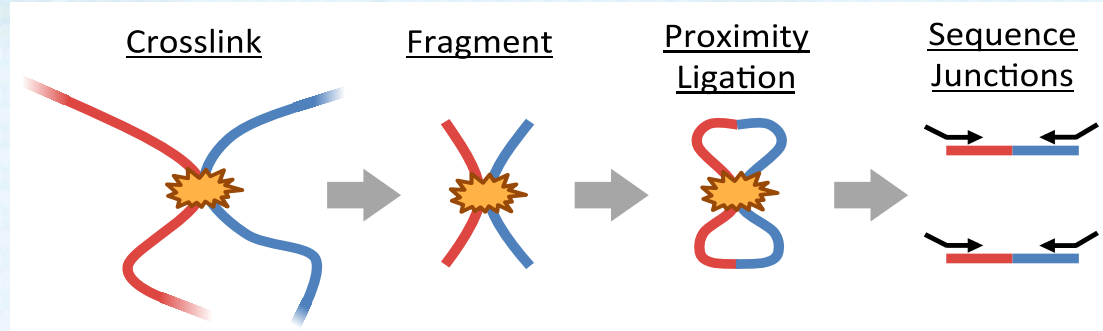
# Proximity Ligation captures ultra-long genomic contiguity



- Proximity in 3D is correlated with genomic distance
- Can be used to:
  - Scaffold and phase a genome of any size
  - Find rearrangements

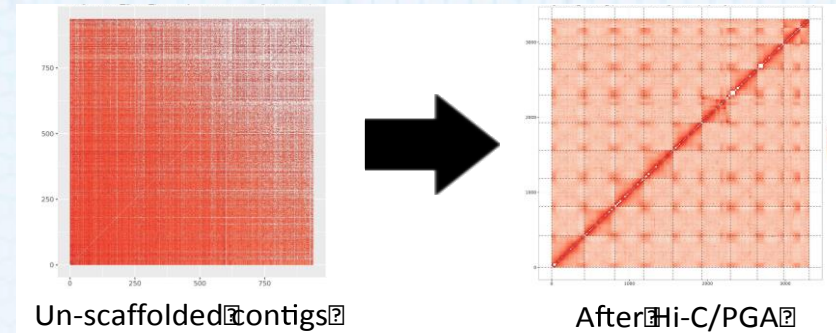


# Using Hi-C to assemble chromosome-scale genome scaffolds




## Proximity-Guided Assembly™ :


- Clustering contigs into chromosome groups
- Ordering and orienting the contigs in scaffolds




# Hi-C becomes a routine tool in eukaryotic genome assembly

- Human (Burton *et al.* 2013 *Nature Biotech*)
- Goat (Bickhart *et al.* 2017 *Nature Genetics*)
- Stickleback (Peichel *et al.* 2017 *Heredity*)
- Amaranth (Lightfoot *et al.* 2017 *BMC Biology*)
- Firefly (Fallon *et al.* 2017 *BioRxiv*)
- Black raspberry (Jibran *et al.* 2018 *Hort. Res.*)
- Clownfish (Lehmann *et al.* 2018 *BioRxiv*)
- Sugar beet (Funk *et al.* 2018 *Plant J*)
- Malaria Mosquito (Ghurye *et al.* 2018 *BioRxiv*)
- *Cannabis* (McKernan *et al.* 2018 *OSF*)
- *Cannabis* (Grassa *et al.* 2018 *BioRxiv*)
- *E. festucae* (Winter *et al.* 2018 *PLoS Genetics*)
- Honeybee (Wallberg *et al.* 2018 *BioRxiv*)
- Aphid (Chen *et al.* 2018 *BioRxiv*)
- *T. inflatum* (Olarte, *et al.* 2019 *BMC Genomics*)
- Bee mites (Techer *et al.* 2019 *BioRxiv*)
- ...more from other labs...

**Michelle Vierra**  
@the\_mvierra



Goat genome is the "greatest of all time" (G.O.A.T...get it?) 😂@PacBio #genomepuns [genome.gov/27567880/](https://genome.gov/27567880/)




**SCIENCE**


## The Game-Changing Technique That Cracked the Zika-Mosquito Genome


"Hi-C" will make it much easier and cheaper to assemble all of an organism's genetic material from scratch.


**ED YONG** MAR 29, 2017



**MORE STORIES**

**A Troubling Discovery in the Deepest Ocean Trenches**  
ED YONG 

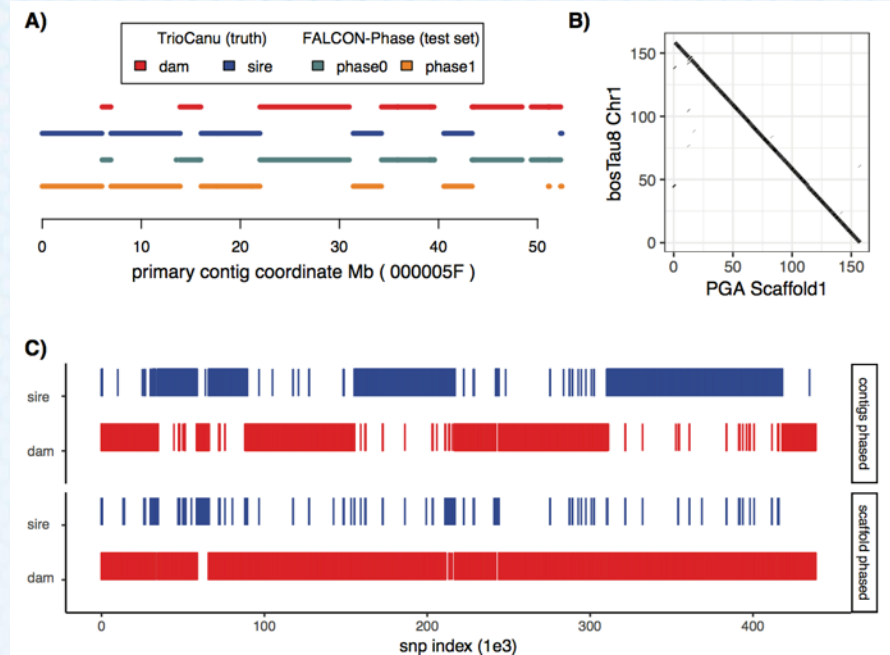
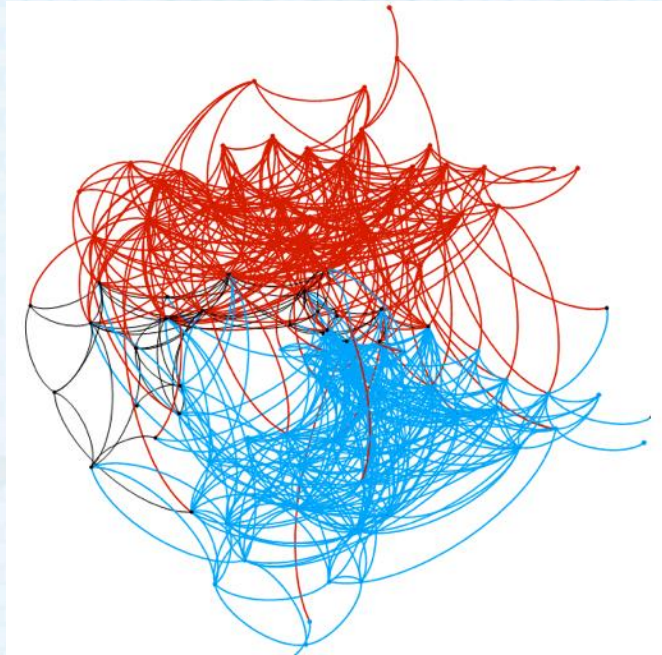
**Grieving Parents Are Turning to Posthumous IVF**  
SHIRA RUBIN AND UNDAK 

**NASA Is Rushing to the Moon**  
MARINA KOREN 

**The Mystery of**



# FALCON-Phase: Combining SMRT and Hi-C data to generate fully phased genome assemblies.

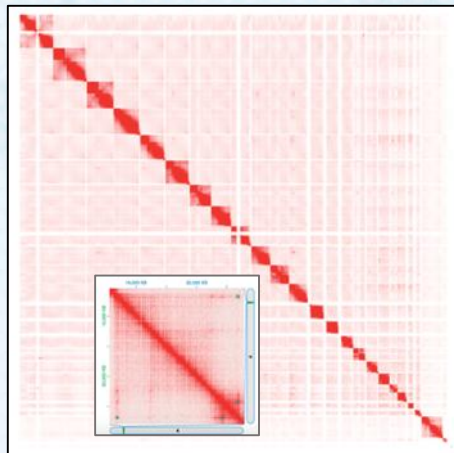




# Proximo™ yields high-res karyotype/SV data

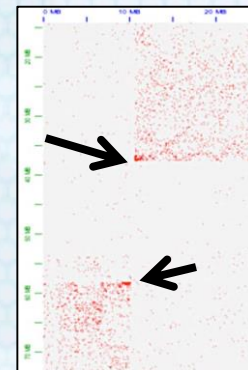
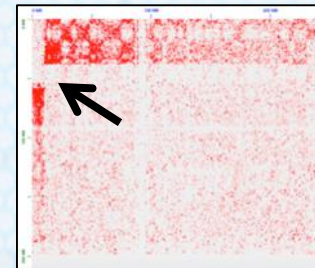
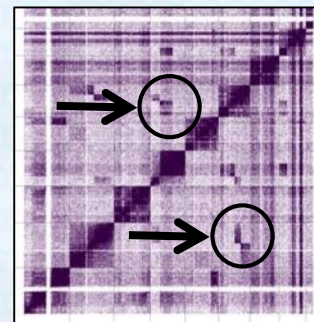
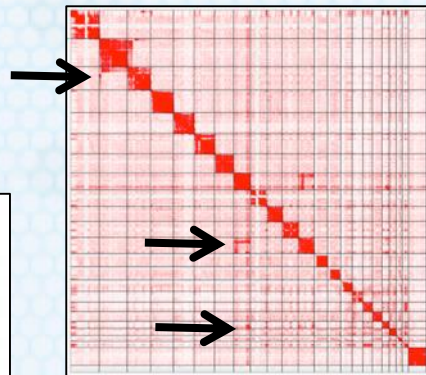
- Rapid
- Scalable
- No special machine
- Works on any sample type

**Normal**

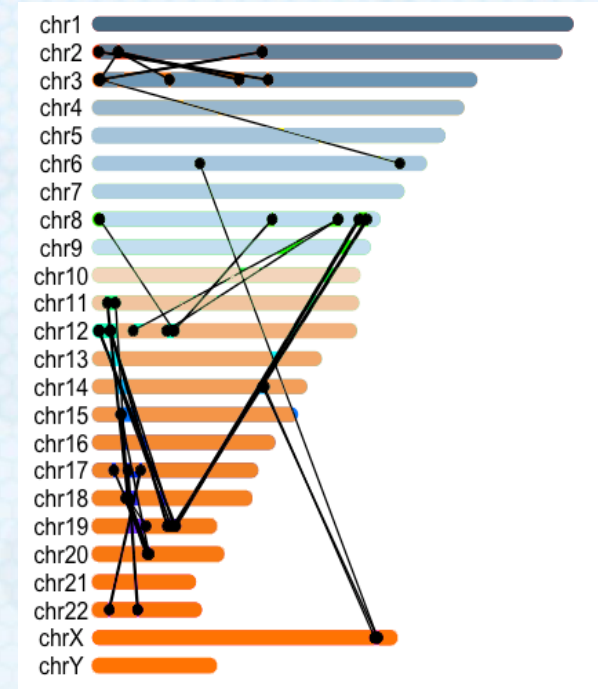
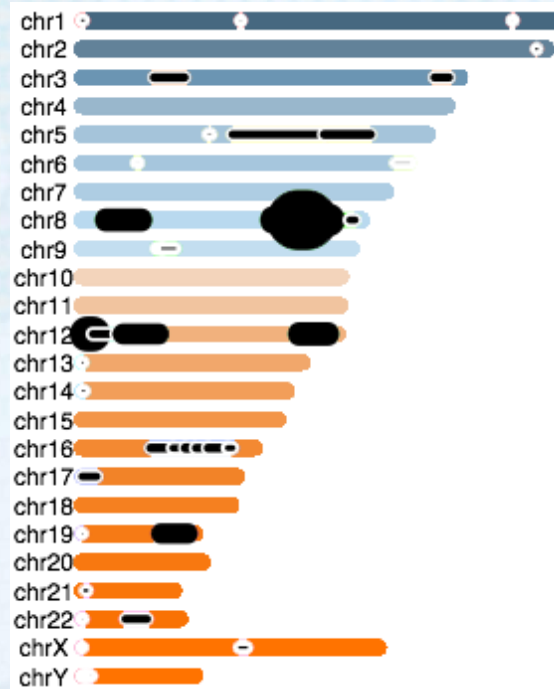
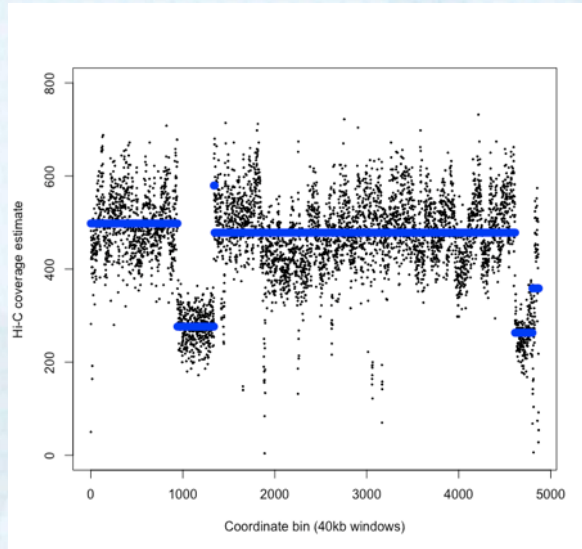


Yardimci and Noble, Genome Biol, 2017

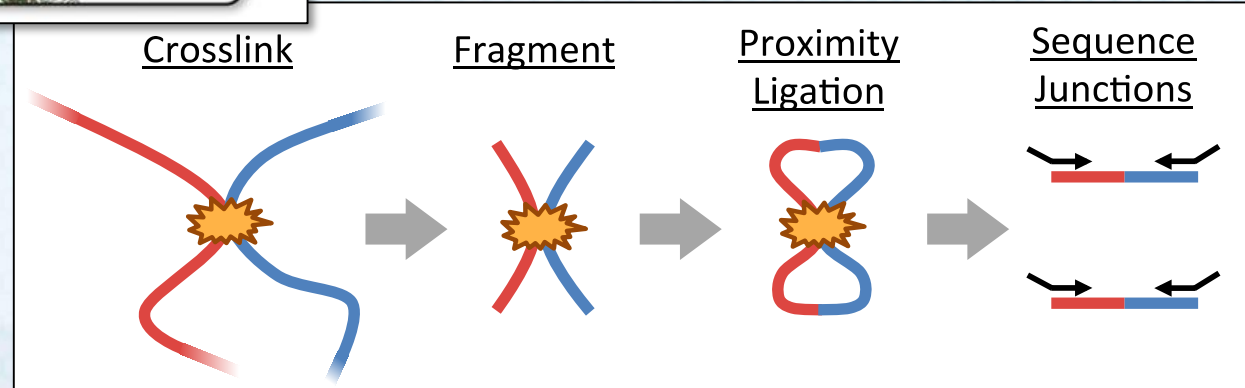
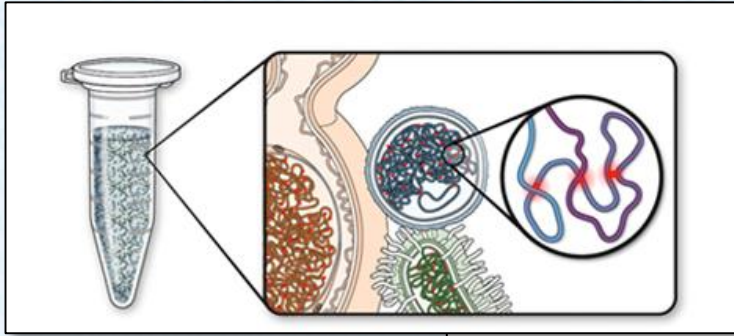
**Cancer**



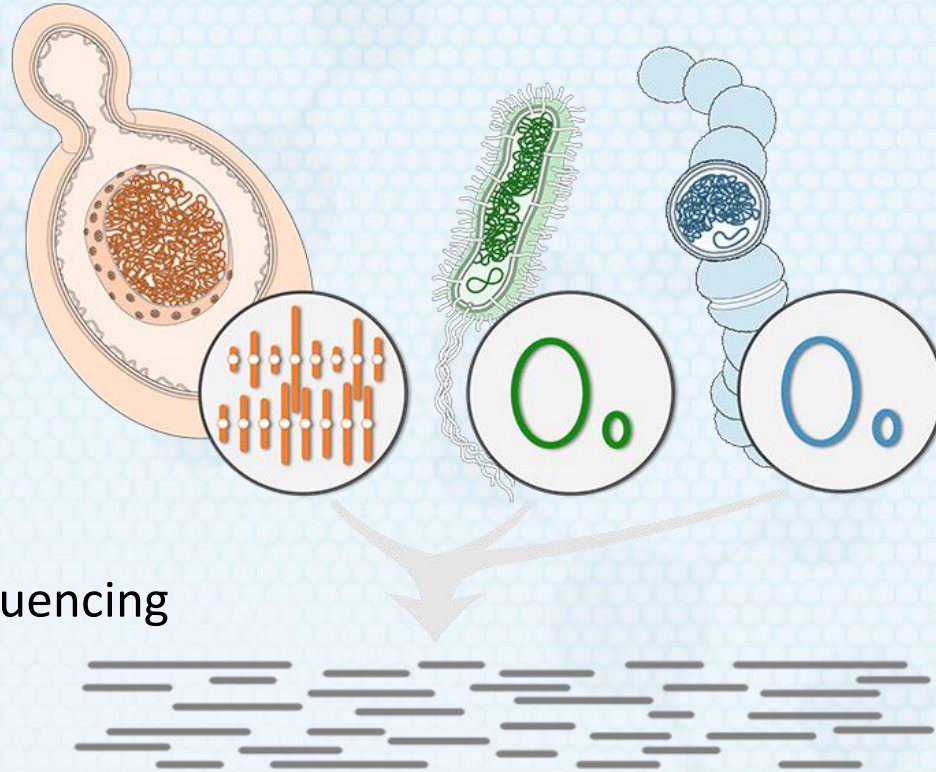
# Simultaneous CNV and SV delineation in AML cells



**Any sequences that interact by Proximity must have originated from the same cell**

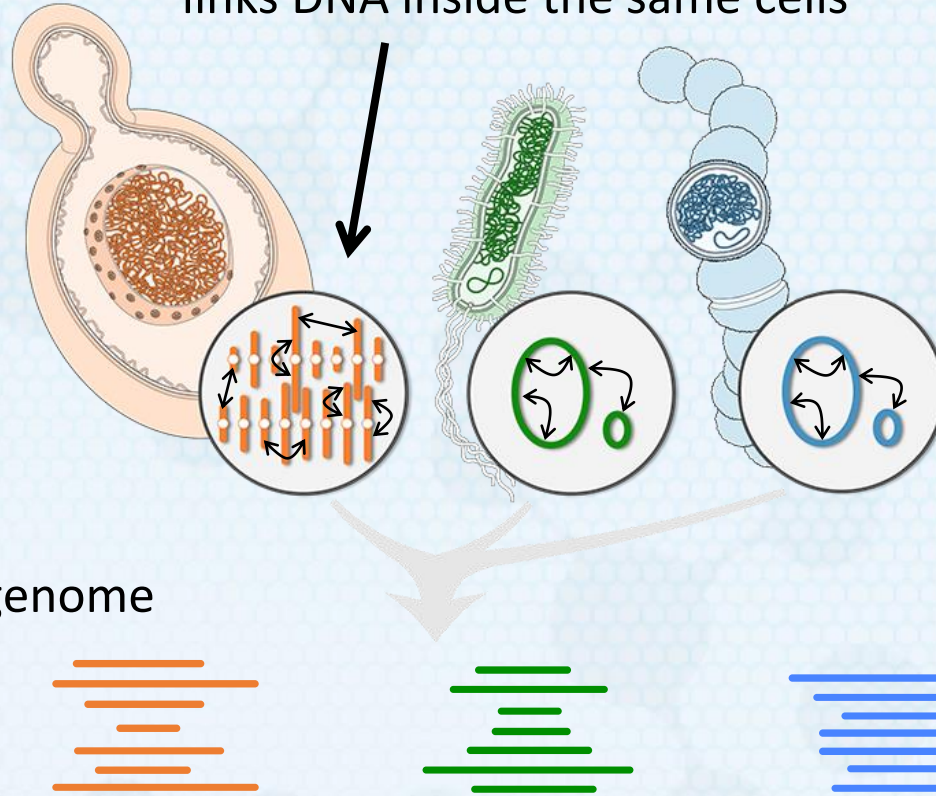






Shotgun sequencing

Proximity ligation chemically  
links DNA inside the same cells

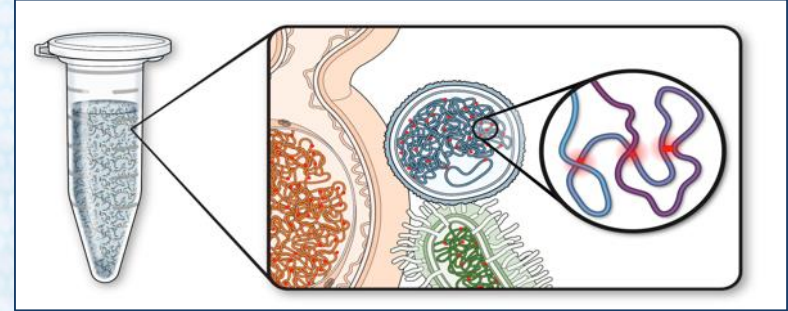


Connects metagenome  
sequences

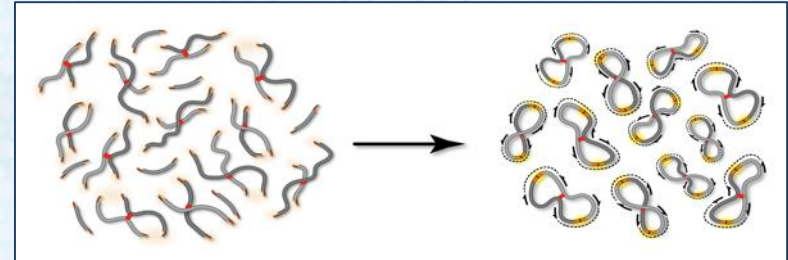


# Proximity-Guided Metagenome Assembly (ProxiMeta™)

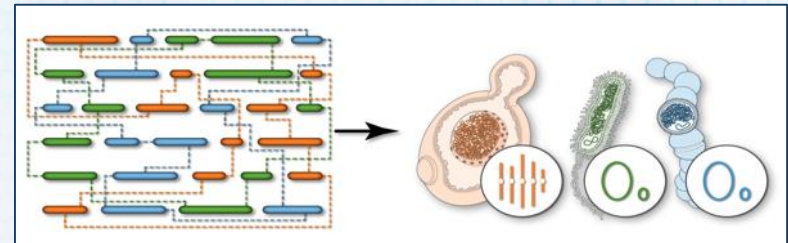
Crosslink intact cells to capture intra-  
cellular interactions



Isolate and sequence crosslinked  
junctions

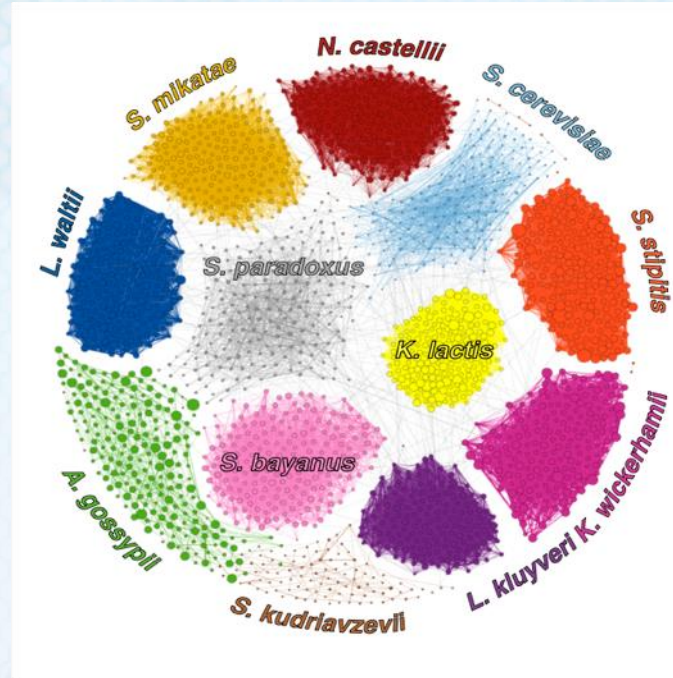


Use proximity connections to  
deconvolute metagenome





# Reference-quality pro- and eu- karyotic genomes from mixed populations



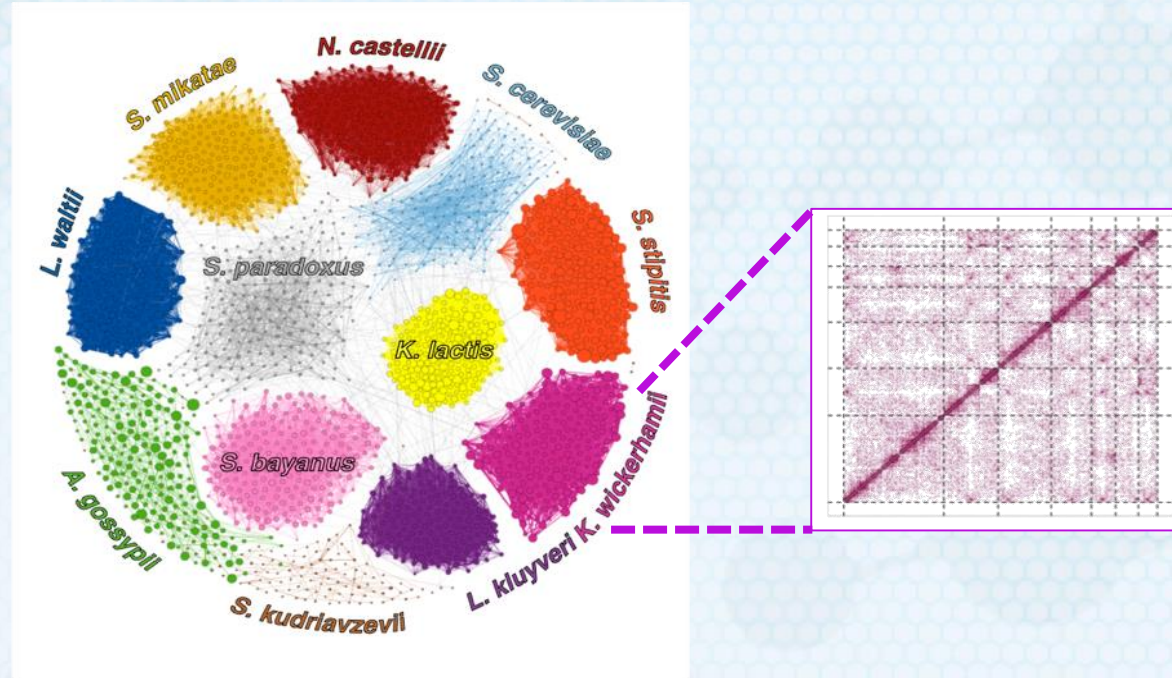
Draft assembly:

Size = 135.2 Mbp

Contig N50 = 17.3 Kb

**Error rate <1%**

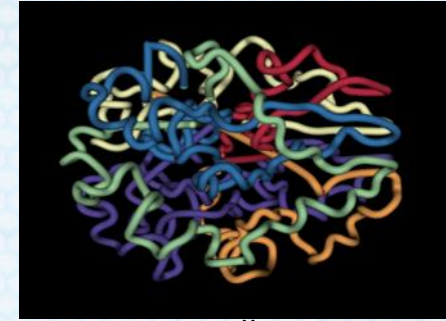
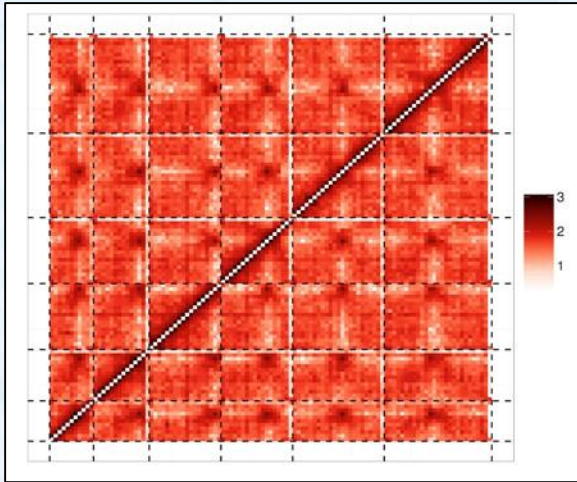
# Reference-quality pro- and eu- karyotic genomes from mixed populations



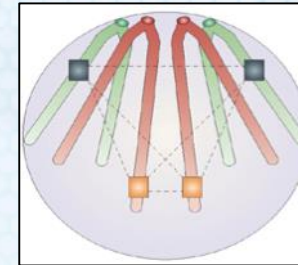


# 3D modeling of genomes directly from mixed populations

*Kluyveromyces lactis*



Nelle Varoquaux

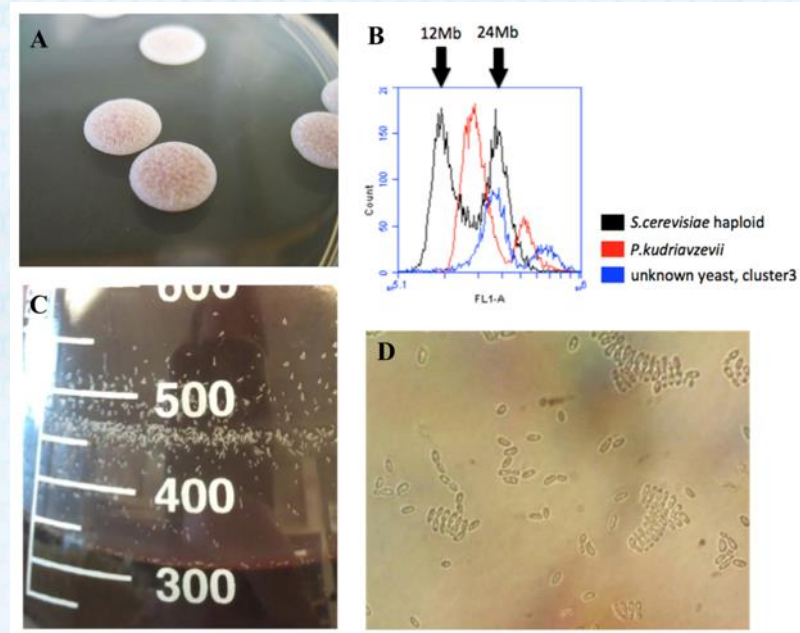
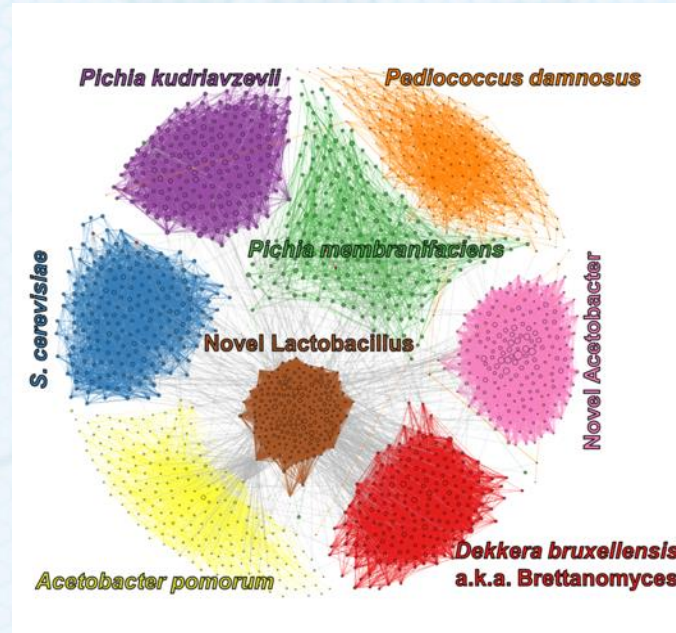


Barzel and Kupiec, 2008

\*Dark spots in the middle of each chromosome are centromeric regions.



# Assembly of a hybrid yeast from a beer metagenome



# Assembly of a hybrid yeast from a beer metagenome

*Pichia kudriavzevii*

В «диком» пиве нашли новый вид дрожжей



скопом  
bioRxiv:150722

ские ученые обнаружили в пиве, ферменты микроорганизмов окружающей среды и дрожжей. Отчет о работе доступен в архиве, также о ней пишет Science.

*Ekkeria bruxellensis*  
k.a. *Brettanomyces*

WILDES BIER

## Neue Mikrobenart in Brauerei entdeckt

Eigentlich wollten US-Biologen in einer Brauerei bloß das Erbgut exotischer Hefepilze analysieren. Doch in einer Probe entdeckten sie etwas Unerwartetes.

von Robert Gast



© Stock / Yuri Arcurs (Ausschütt)

In dem offenen Fass am Rande der Brauerei in Seattle ging etwas Ungewöhnliches vor, das erkannten die Wissenschaftler auf den ersten Blick. Die Genetiker der University of Washington wollten bei ihrem Besuch Proben von Hefepilzen



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Open fermentation vessels at Pilsner Unquell brewery in Pilsen, Czech Republic. apothecary stock photo

**Microbe new to science found in self-fermented beer**

By Aleszu Bajak | Jul. 28, 2017, 1:30 PM

In May 2014, a group of scientists took a field trip to a small brewery in an old warehouse in



La mayoría de cervezas se fermentan con la ayuda de levaduras como *Saccharomyces cerevisiae*.

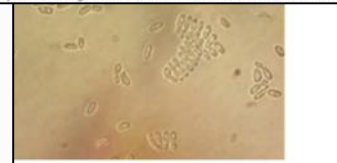
CIENCIA / BEBIDAS ALCOHÓLICAS

## Esta es la cerveza artesana que ha permitido descubrir un nuevo microbio

Un equipo de la Universidad de Seattle encuentra por casualidad una levadura desconocida hasta la fecha por la comunidad científica.  
1 agosto, 2017 - 02:25

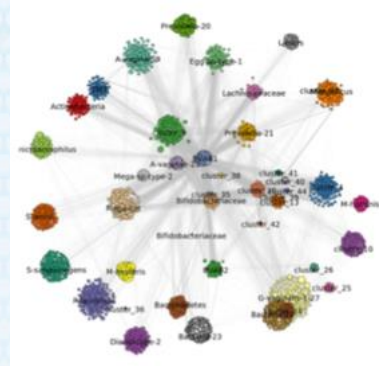
EN: CERVEZA MICROBIOLOGÍA Y PARASITOLOGÍA SEATTLE

José Andrés Gómez





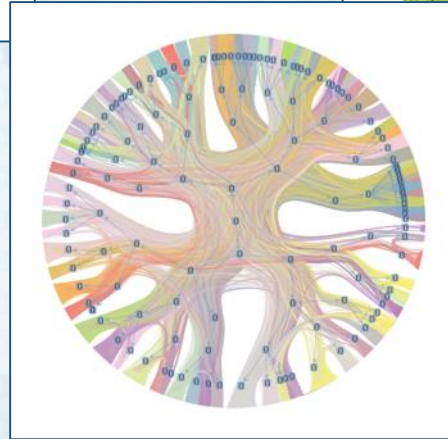
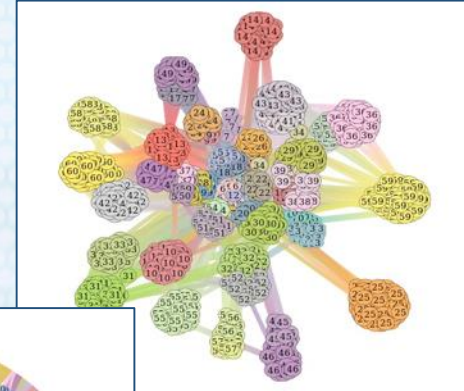
# New genomes and strains from a bacterial vaginosis sample



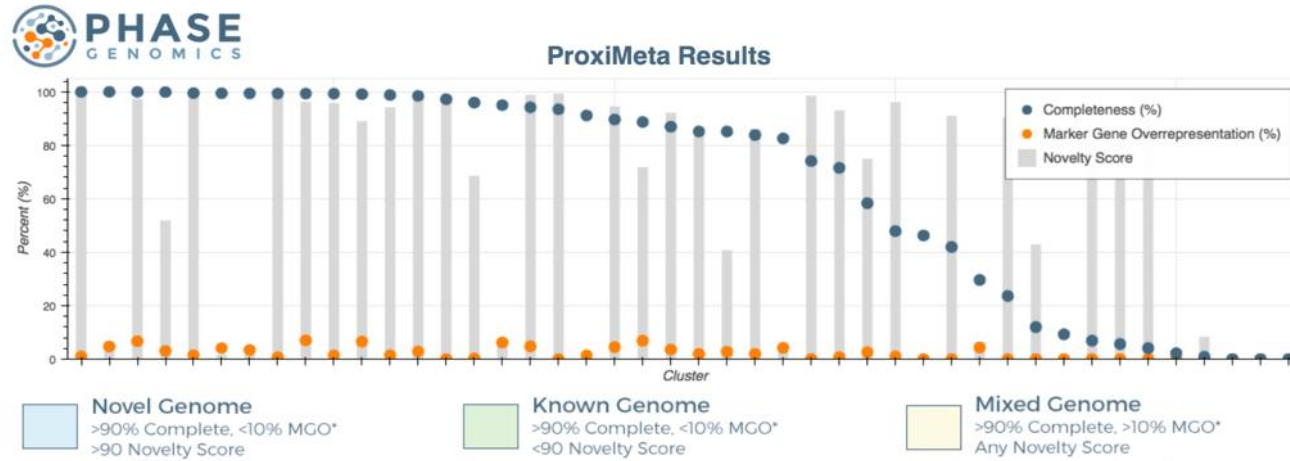
- ProxiMeta clustering of assembly containing all three read sets (N50  $\approx$  17 kb) yielded >20 high quality draft genomes with >95% core gene groups and N50 >20 kb.
- Least abundant species yielding a high quality genome was represented at 0.2% RA in the combined read set.
- Eight of the high quality genomes came from novel and/or previously unsequenced species (starred).
- PGA successfully segregated some strains that differed vastly in protein sequence identity.

	SPECIES	LENGTH (Mb)	N50 (Kb)
1	Gardnerella vaginalis	2.4	7.9
2	Gardnerella vaginalis	0.9	4.3
3	Atopobium vaginae	1.6	22.6
4	* Eggerthella sp. type 1	2.1	39.1
5	Prevotella	0.8	9.3
6	Peptoniphilus lacrimalis	1.7	21.3
7	Gardnerella vaginalis	1.1	8.5
8	Gardnerella vaginalis	1.1	9.5
9	Gardnerella vaginalis	0.6	6.1
10	Prevotella	2.7	38.7
11	Megasphaera sp. type 1	3.6	72.5
12	Gardnerella vaginalis	1.1	12.9
13	* Sneathia sanguinegens	1.0	9.9
14	Gardnerella vaginalis	1.7	35.0
15	Atopobium vaginae	1.6	33.7
16	* BVAB2	1.6	24.4
17	Gardnerella vaginalis	0.6	7.9
18	Dialister microaerophilus	1.2	24.5
19	* Sneathia amnii	1.1	15.2
20	* Dialister sp. type 2	1.5	33.5
21	BVAB3	1.6	48.0
22	Mycoplasma hominis	0.4	7.9
23	* Prevotella	2.2	67.7
24	* BVAB1	1.7	109.5
25	* BVAB4	1.5	40.6
26	Atopobium vaginae	1.6	144.8
27	* Prevotella	2.4	94.1
28	Gardnerella vaginalis	1.1	23.1
29	* TM7	1.2	34.1
30	Lactobacillus iners	1.4	64.8
31	Mobiluncus mulieris	2.1	118.2
32	Gardnerella vaginalis	0.1	10.8
33	Megasphaera sp. type 1	0.0	3.9
34	Gardnerella vaginalis	0.0	5.0
35	Gardnerella vaginalis	0.2	35.6
36	Gardnerella vaginalis	0.0	10.6
37	Atopobium vaginae	0.1	59.5
38	Gardnerella vaginalis	0.0	25.0





# Novel, high-completeness genomes from diverse samples





\*Marker Gene Overrepresentation

Cluster ID	Top Reference	Completeness (%)	Marker Gene Overrepresentation (%)	Novelty Score	Abundance (%)	GC (%)	Genome Size	Num Contigs	Contig N50
level_9.cluster.4	<i>p__Bacteroidetes</i>	100.00	1.03	99.20	2.17	47.84	6,051,721	272	88,894
level_8.cluster.6	<i>Leucobacter</i> sp. UCD THU	100.00	4.78	3.36	2.27	69.14	4,017,073	256	26,530
level_12.cluster.13	<i>p__Proteobacteria</i>	100.00	6.71	97.23	3.91	64.60	4,187,026	133	97,353
level_8.cluster.0	<i>Cupriavidus</i> sp. HPC L	99.89	3.06	51.67	2.20	67.66	5,806,781	109	167,085
level_11.cluster.9	<i>c__Betaproteobacteria</i>	99.53	1.45	98.40	2.31	49.42	3,226,972	303	26,412

In collaboration with Herminia Loza-Tavera, Ayixon Sanchez-Reyez



## Degradation of recalcitrant polyurethane and xenobiotic additives by a selected landfill microbial community and its biodegradative potential revealed by proximity ligation-based metagenomic analysis

Itzel Gaytán,  Ayixon Sánchez-Reyes, Manuel Burelo, Martín Vargas-Suárez, Ivan Liachko, Maximilian Press, Shawn Sullivan, Javier Cruz-Gómez,  Herminia Loza-Tavera

doi: <https://doi.org/10.1101/760637>

This article is a preprint and has not been certified by peer review [what does this mean?].

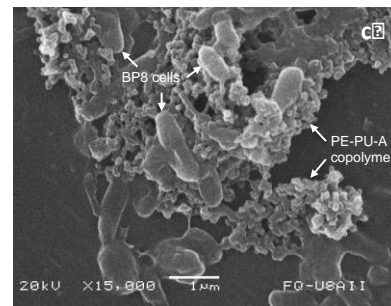
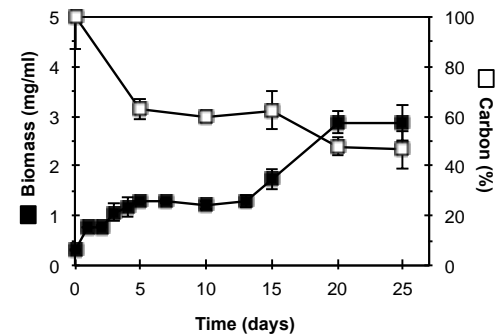
Abstract

Full Text

Info/History

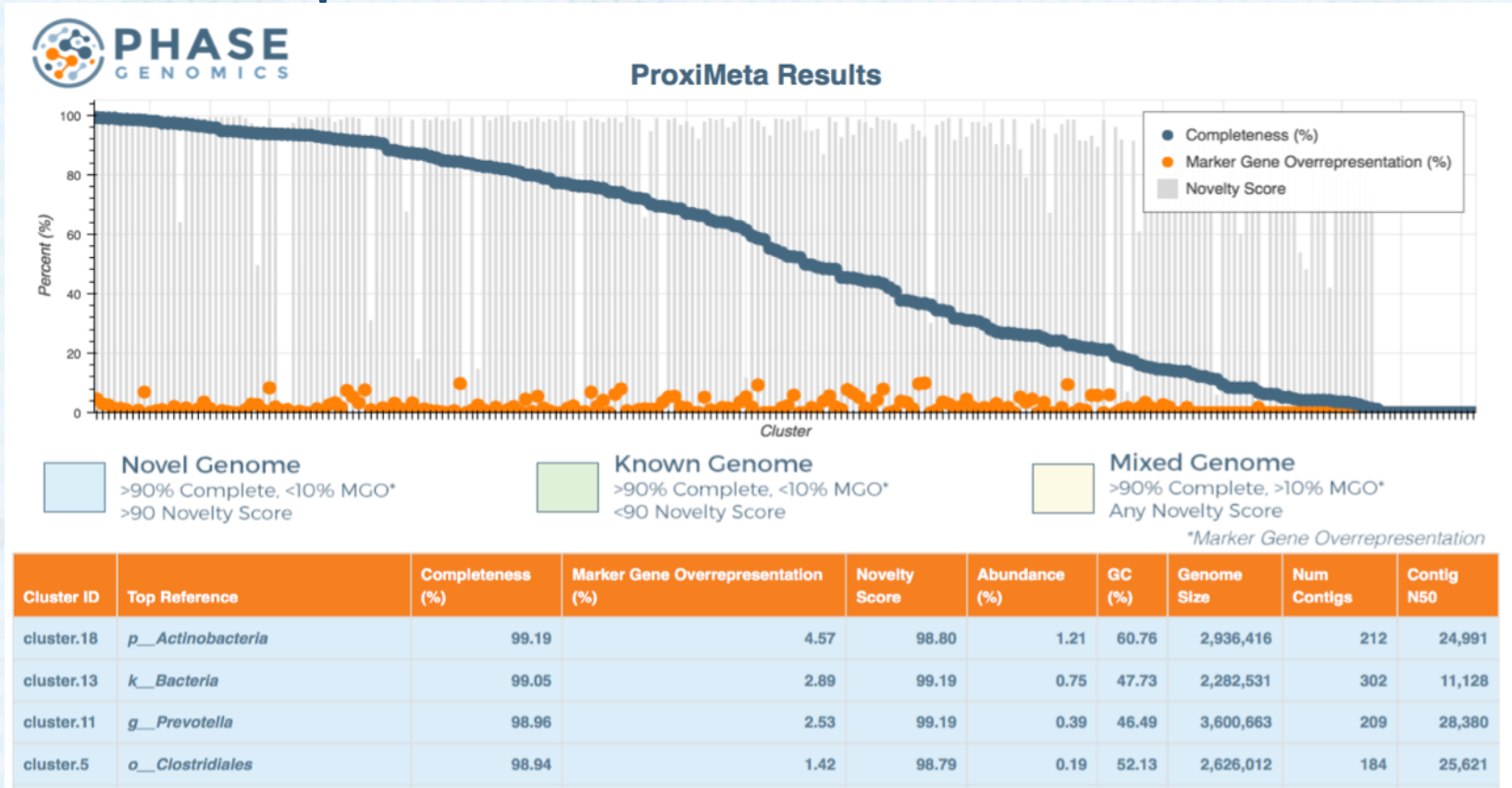
Metrics

 Preview PDF





# High numbers of high-quality, novel genomes directly from rumen samples



# High numbers of high-quality, novel genomes directly from rumen samples



ProxiMeta



FOOD FOR THOUGHT

## Mysteries of the Moo-crobiome: Could Tweaking Cow Gut Bugs Improve Beef?

March 6, 2018 · 8:00 AM ET

MENAKA WILHELM



Altmetric: 565

[More detail >>](#)

Article | [OPEN](#)

## Assembly of 913 microbial genomes from metagenomic sequencing of the cow rumen

Robert D. Stewart, Marc D. Auffret, Amanda Warr, Andrew H. Wiser, Maximilian O. Press, Kyle W. Langford, Ivan Liachko, Timothy J. Snelling, Richard J. Dewhurst, Alan W. Walker, Rainer Roehe & Mick Watson

*Nature Communications* 9,  
Article number: 870 (2018)

Received: 26 October 2017  
Accepted: 05 February 2018

cluster.11 *g\_\_Prevotella*  
cluster.5 *o\_\_Clostridiales*

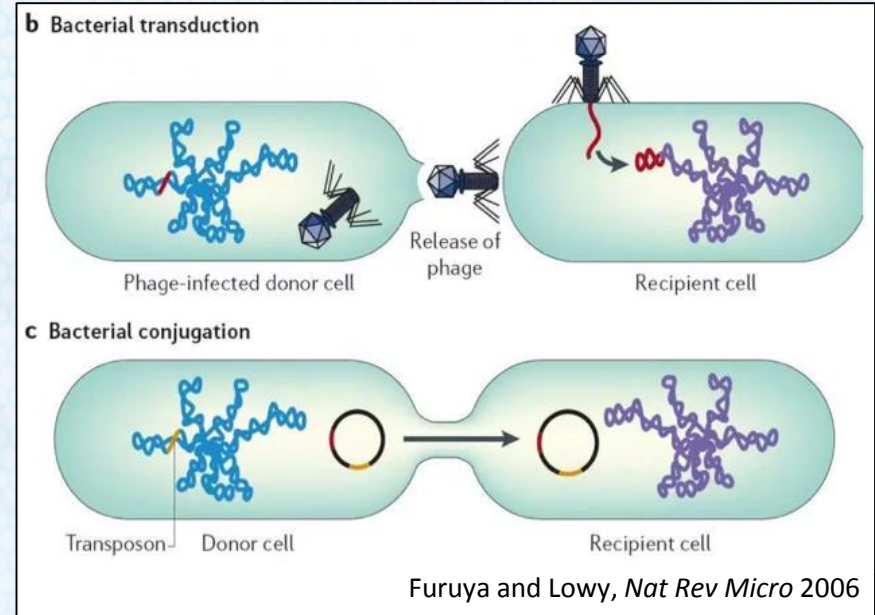
98.96  
98.94

2.53  
1.42

*Marker Gene Overrepresentation					
Novelty Score	Abundance (%)	GC (%)	Genome Size	Num Contigs	Contig N50
98.80	1.21	60.76	2,936,416	212	24,991
99.19	0.75	47.73	2,282,531	302	11,128
99.19	0.39	46.49	3,600,663	209	28,380
98.79	0.19	52.13	2,626,012	184	25,621

# Plasmids/viruses are key players in the microbiome

- Plasmids/Phage transmit AMR (Anti-microbial resistance)
- Plasmids often transmit pathogenic/toxic genes (ex. Anthrax)
- Virtually impossible to connect AMR and mobile elements with host strains using normal NGS
- Need a method that can directly link plasmids/viruses to hosts.



## ABSTRACT

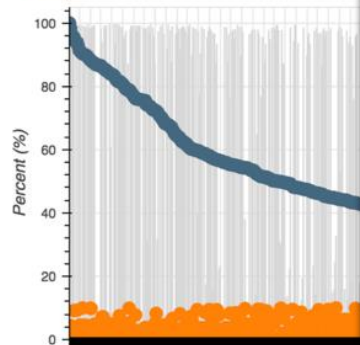
[Go to:](#) ☒

In order to cause the disease anthrax, *Bacillus anthracis* requires two plasmids, pX01 and pX02, which carry toxin and capsule genes,

Luna *et. al.*, *J Clin Microbiol.* 2006 Jul; 44(7): 2367–2377

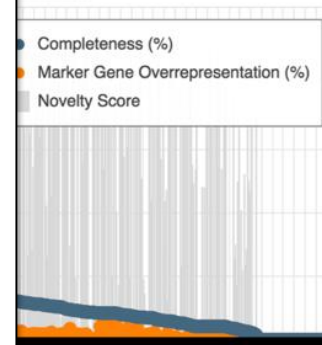
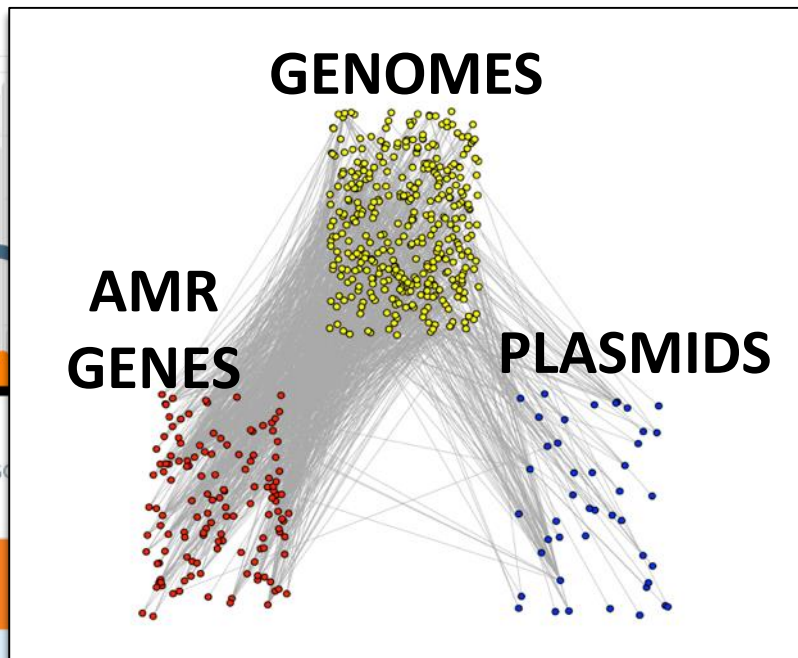


# Highly complex wastewater community



Novel Genome  
>80% Complete, <10% MGO  
>90 Novelty Score

Cluster ID	Top Reference	Abundance (%)	GC (%)	Genome Size	Num Contigs	Contig N50
cluster.1	<i>k_Bacteria</i>	0.14	37.53	2,847,916	86	77,051
cluster.2	<i>Methanobrevibacter_smithii_TS145B.fna</i>	98.40	4.40	4.22	0.06	30.46
cluster.3	<i>p_Firmicutes</i>	96.43	2.22	98.76	0.17	34.55
cluster.4	<i>c_Gammaproteobacteria</i>	95.86	2.26	96.30	0.05	43.38

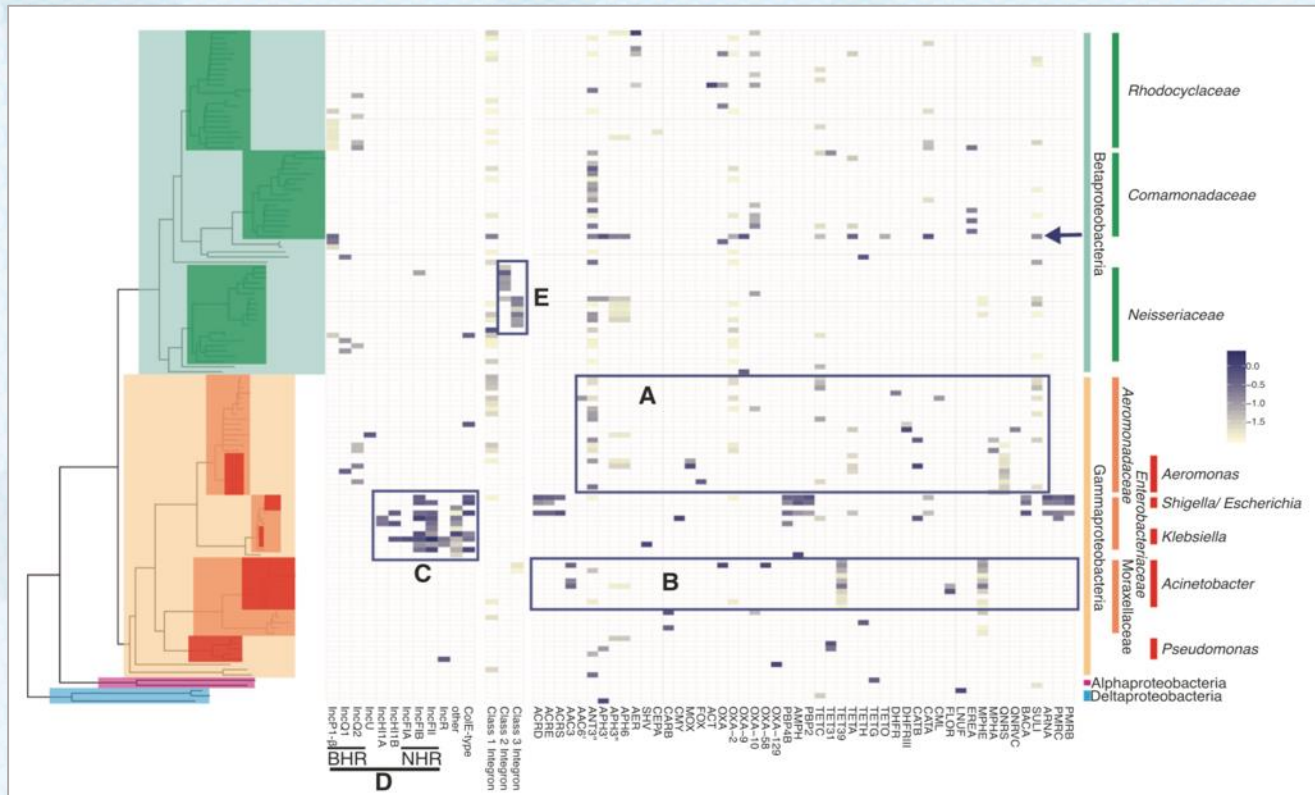


Novel Genome  
Complete, >10% MGO  
Novelty Score  
\*Marker Gene Overrepresentation

# Linking the 'Mobilome' to the Microbiome

Genomes Plasmids Antibiotic resistance genes

Proteobacteria





Phylogenetic tree and heatmap showing the relative abundance of 16S rRNA sequences for various bacterial families across 10 samples (G+, 100mHA, ANT9, ANT8, ANT6, ANT5, TETW, TETD, CENR, ERNG, LUNC, MSRD, MELT, MEFT). The color scale ranges from -0.5 (dark blue) to -1.5 (yellow).

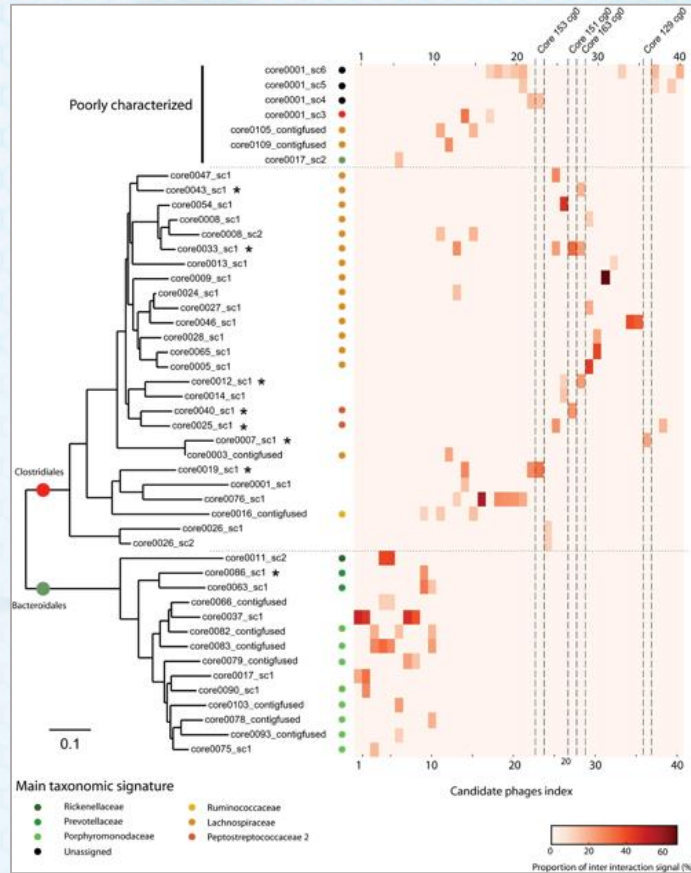
**Bacterial Families:**

- Lachnospiraceae
- Clostridiaceae & Ruminococcaceae
- Eubacteriaceae
- Veillonellaceae & Acidaminococcaceae & Selenomonadaceae
- Streptococcaceae

**Samples:** G+, 100mHA, ANT9, ANT8, ANT6, ANT5, TETW, TETD, CENR, ERNG, LUNC, MSRD, MELT, MEFT.

## 58 integron-host assn

# Tracking viral-host association in metagenomes



Marbouty *et al.*, Science Advances 2017



Stalder *et al.*, ISME J, 2019

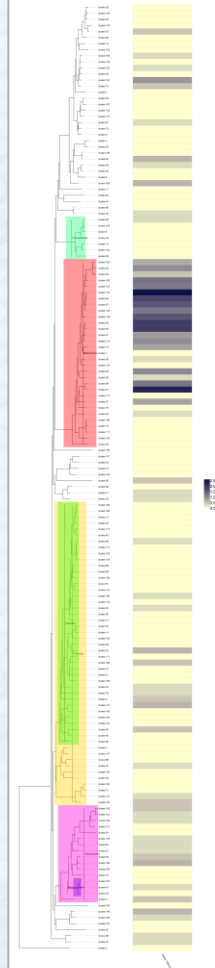


# ex: Where is crAssphage?

Bacterioidetes

Clostridia

Actinobacteria



# Combining long reads and ProxiMeta in a complex microbiome context

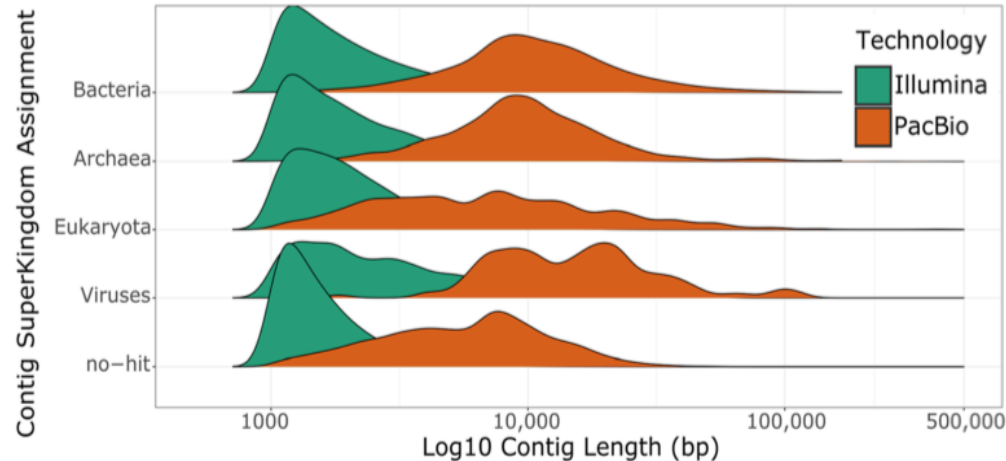
## METHOD

Open Access

Assignment of virus and antimicrobial resistance genes to microbial hosts in a complex microbial community by combined long-read assembly and proximity ligation



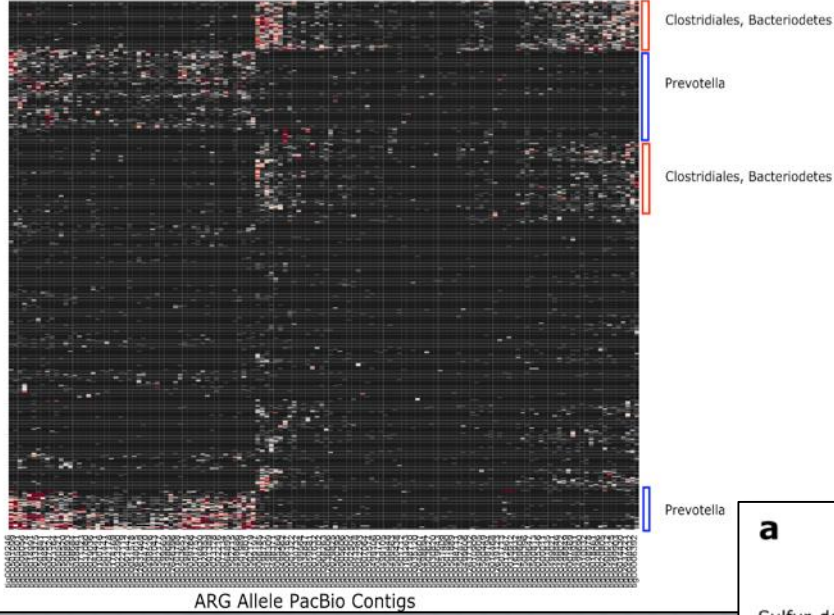
Derek M. Bickhart<sup>1†</sup>, Mick Watson<sup>2†</sup>, Sergey Koren<sup>3†</sup>, Kevin Panke-Buisse<sup>1</sup>, Laura M. Cersosimo<sup>4</sup>, Maximilian O. Press<sup>5</sup>, Curtis P. Van Tassel<sup>6</sup>, Jo Ann S. Van Kessel<sup>7</sup>, Bradd J. Haley<sup>7</sup>, Seon Woo Kim<sup>8</sup>, Garret Suen<sup>9</sup>, Kiranmayee Bakshy<sup>1</sup>, Ivan Liachko<sup>5</sup>, Shawn T. Sullivan<sup>5</sup>, Phillip R. Myer<sup>10</sup>, Jay Ghury<sup>11</sup>, Paul J. Weimer<sup>1,9</sup>, Adam M. Phillippy<sup>3</sup> and Timothy P. L. Smith<sup>12\*</sup>



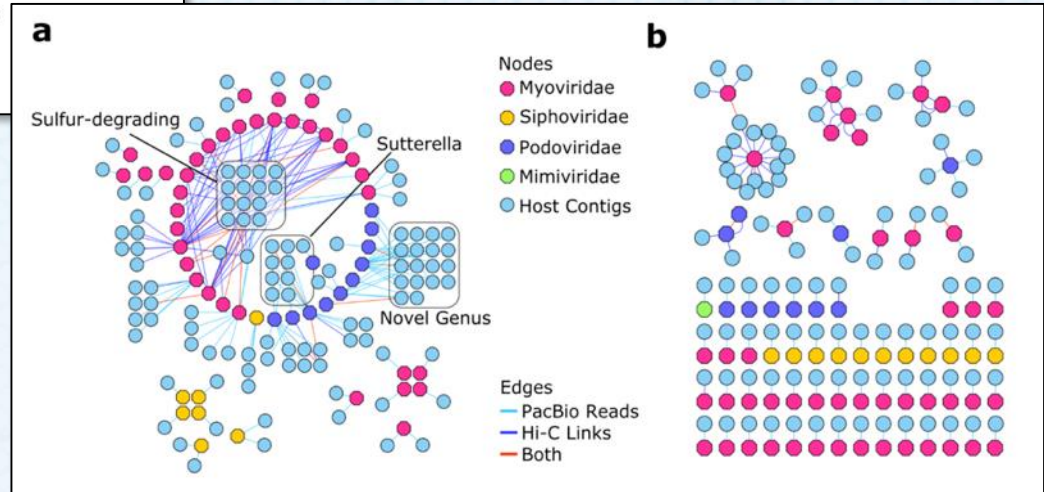


# Connecting ARGs and viruses to their hosts

Proximeta PacBio Assembly Bins



\*188 Novel viruses and host interactions discovered from one rumen sample



**Sneak Peek...**



# The limitations of metagenomic binning

## Human contamination in bacterial genomes has created thousands of spurious proteins

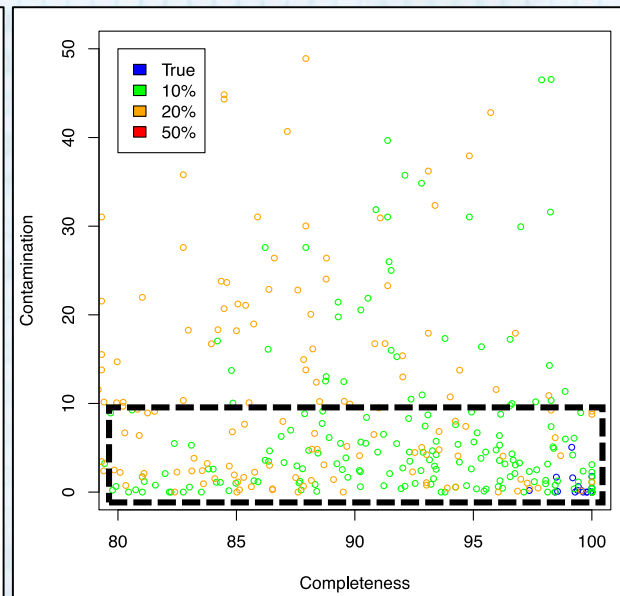
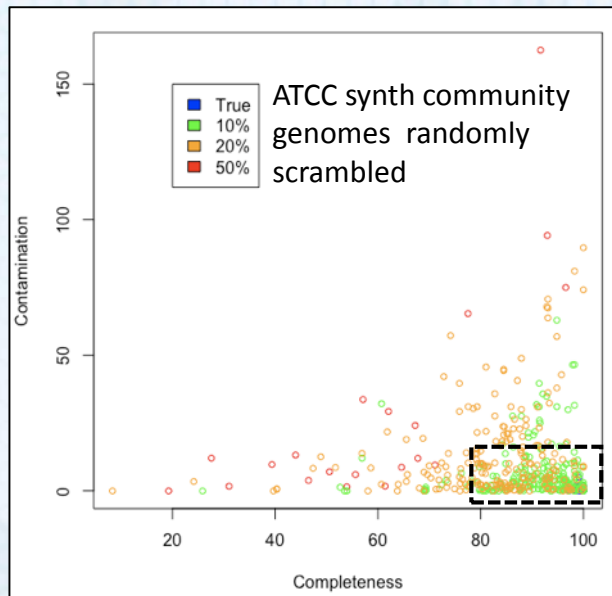
Florian P Breitwieser, Mihaela Perteu, Aleksey Zimin and Steven L Salzberg<sup>1</sup>

## Composite Metagenome-Assembled Genomes Reduce the Quality of Public Genome Repositories

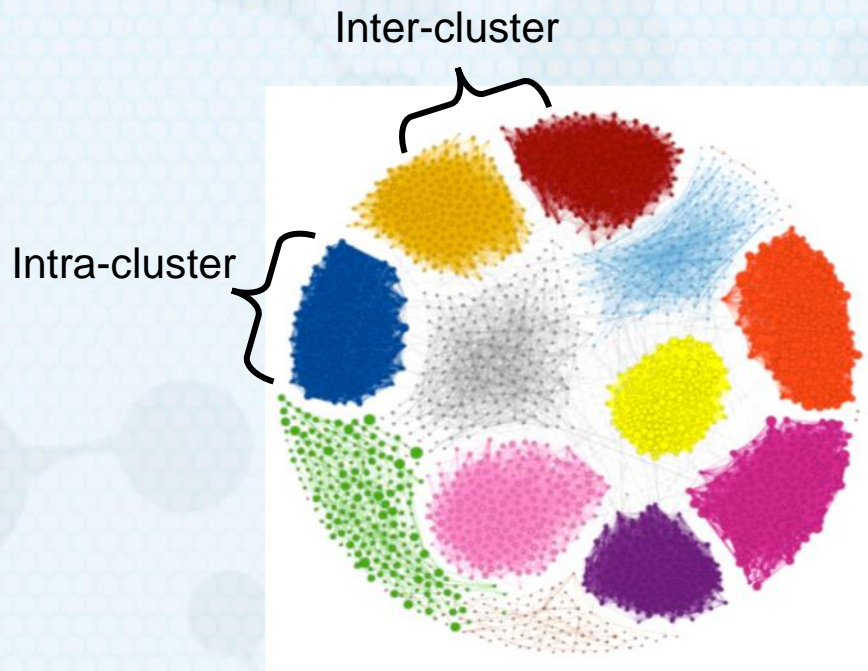
Alon Shaiber,<sup>a</sup> A. Murat Eren<sup>b,c</sup>


CheckM uses core gene content to QC MAGs


Scrambled genomes are often called as High-Quality MAGs



# ProxiMeta data provides a direct orthogonal datatype to QC metagenome bins

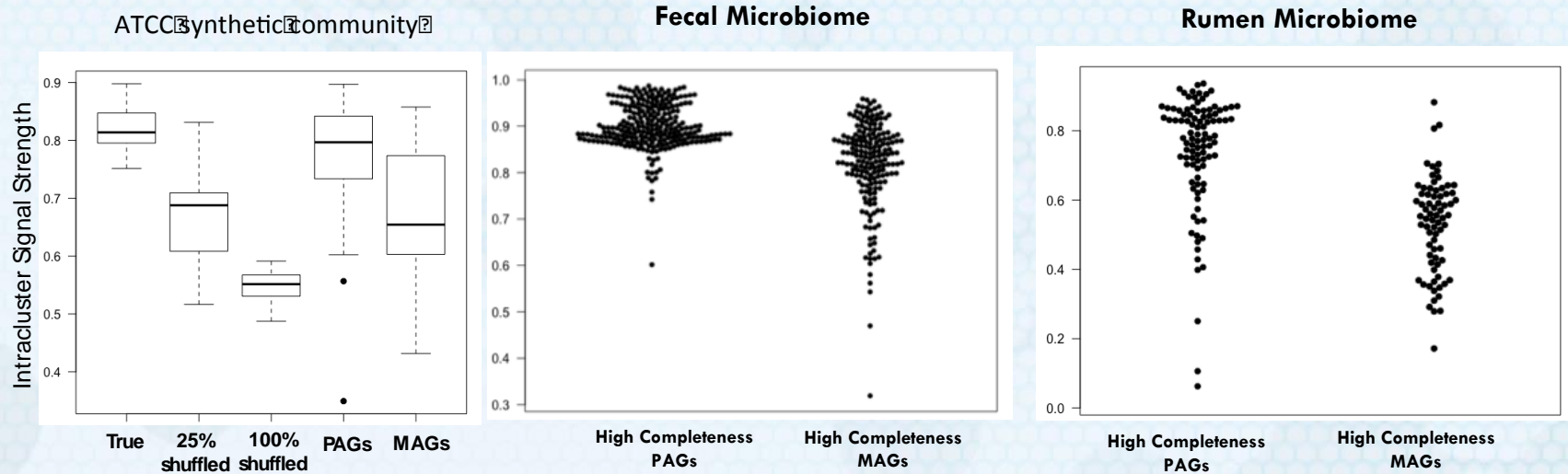


Correctly binned:  $\frac{\text{intra}}{\text{inter}}$  = 

Scrambled:  $\frac{\text{intra}}{\text{inter}}$  = 

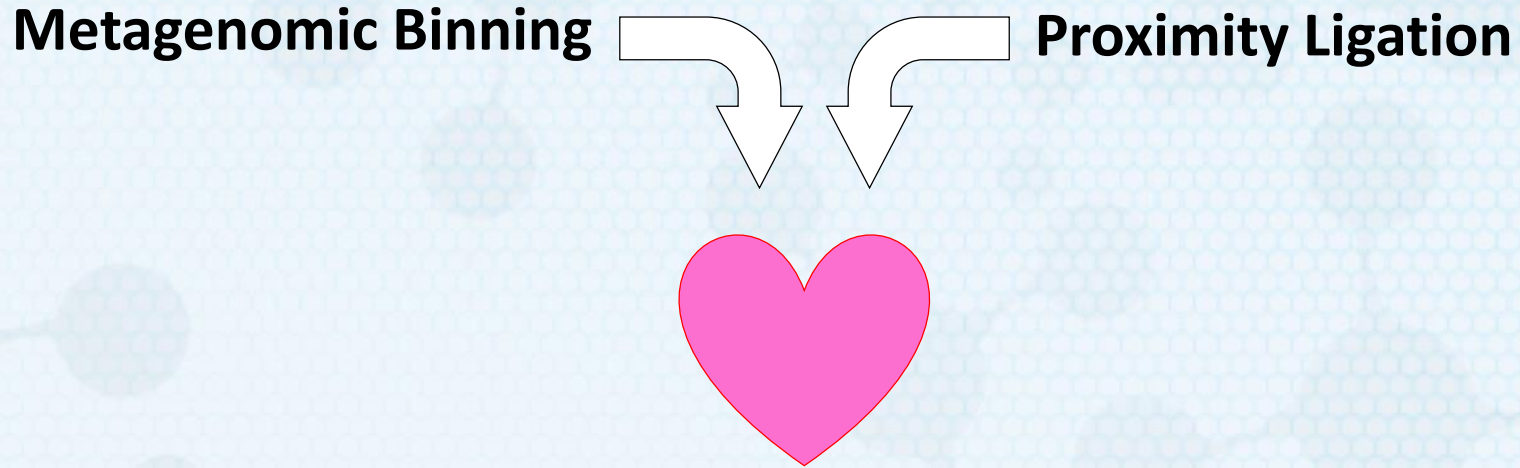


# Proximity-assembled genomes are more accurate than MAGs

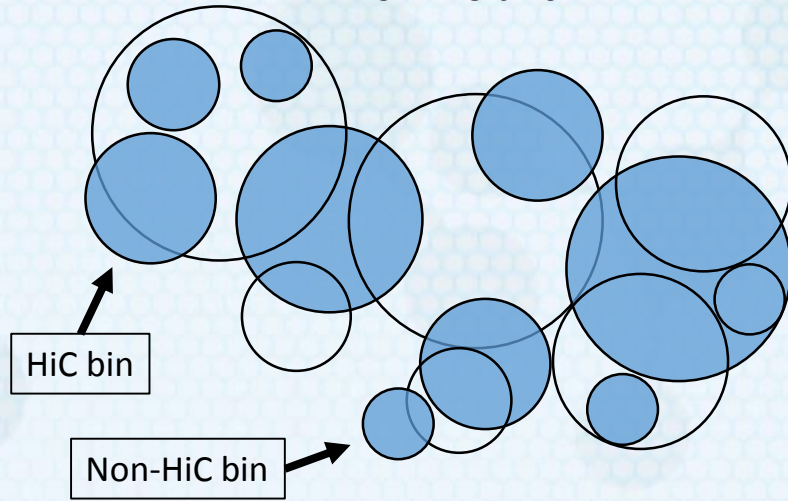


\*High-completeness MAGs demonstrate low intra-cluster enrichment values, indicating high degree of error

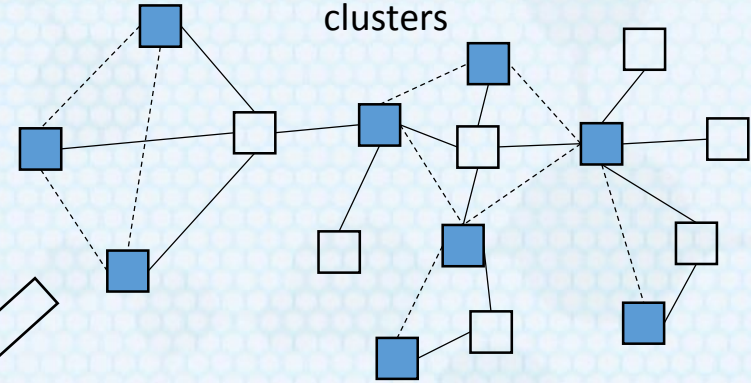
# Can we get the best of both worlds?



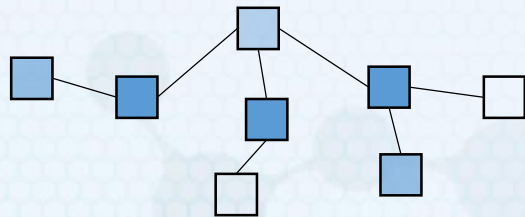
1. Compute overlaps between HiC and non-HiC bins



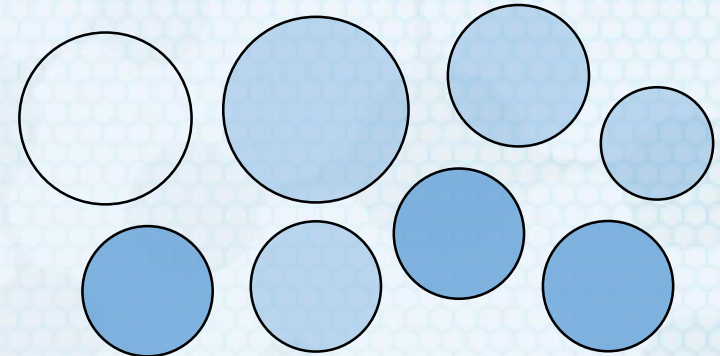
2. Generate connectivity network of contig clusters



3. Supervised algorithm to hierarchically de-convolve network

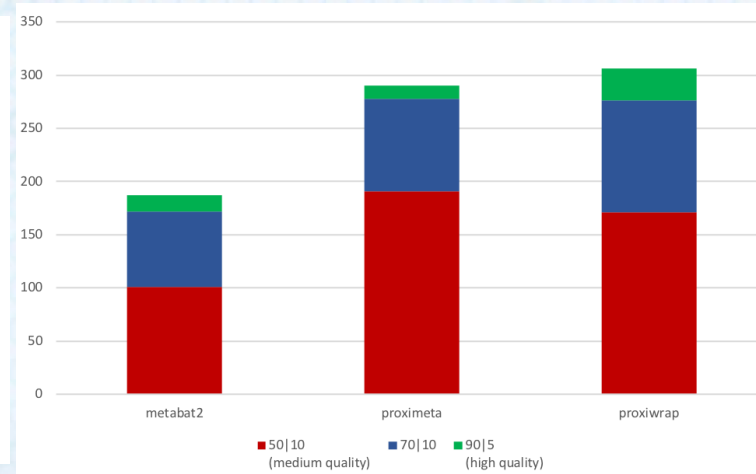
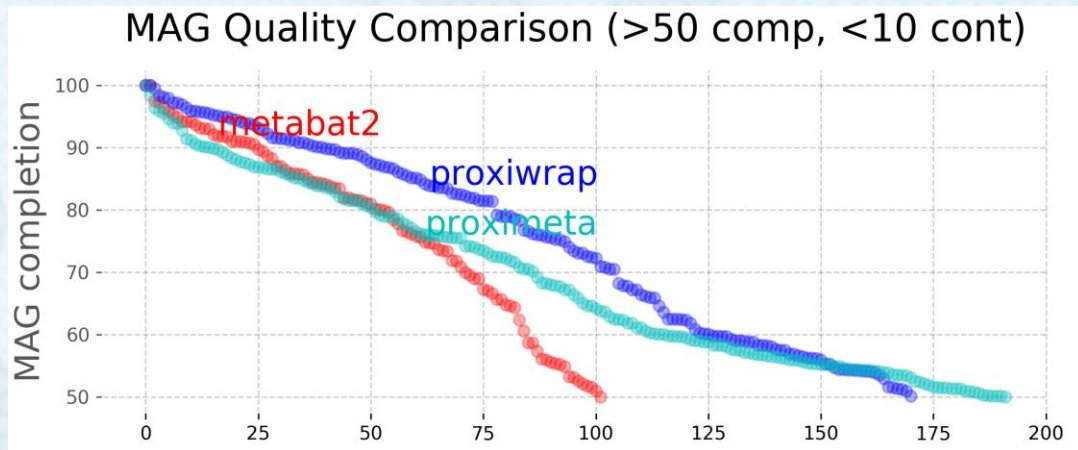


4. Export final set of bins



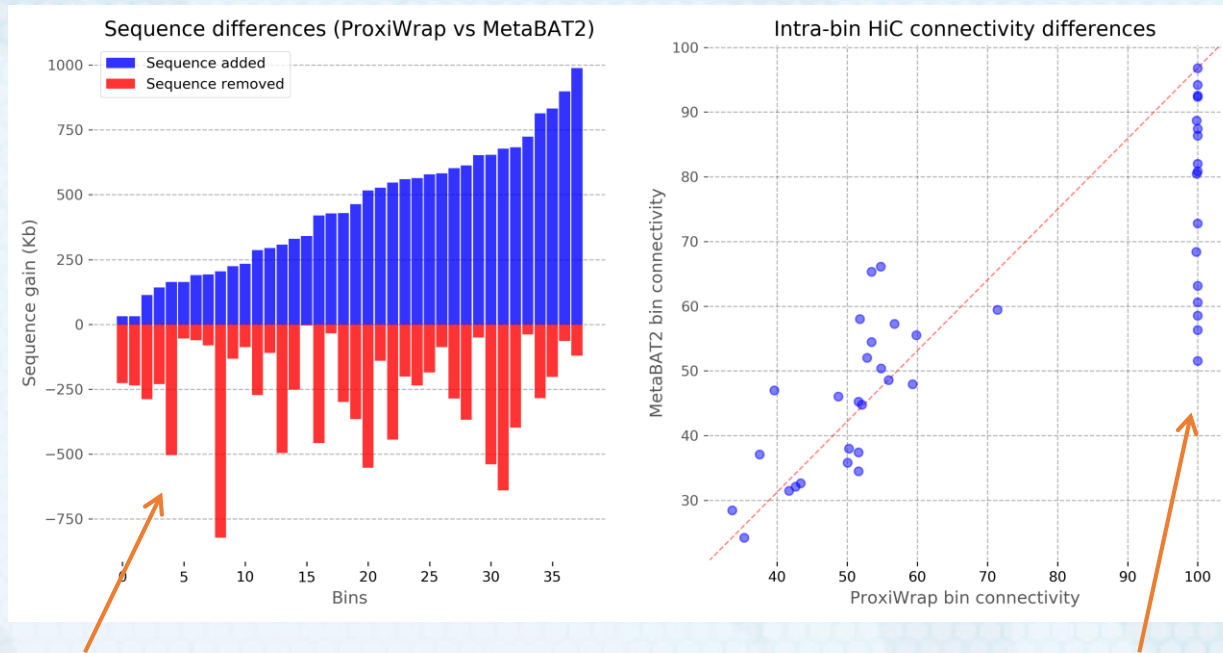


# Applying ProxiWrap™ to highly complex wastewater sample



\*ProxiWrap yields more high quality genomes than metaBAT2 (binning) or ProxiMeta (Hi-C)

# Applying ProxiWrap to highly complex wastewater sample



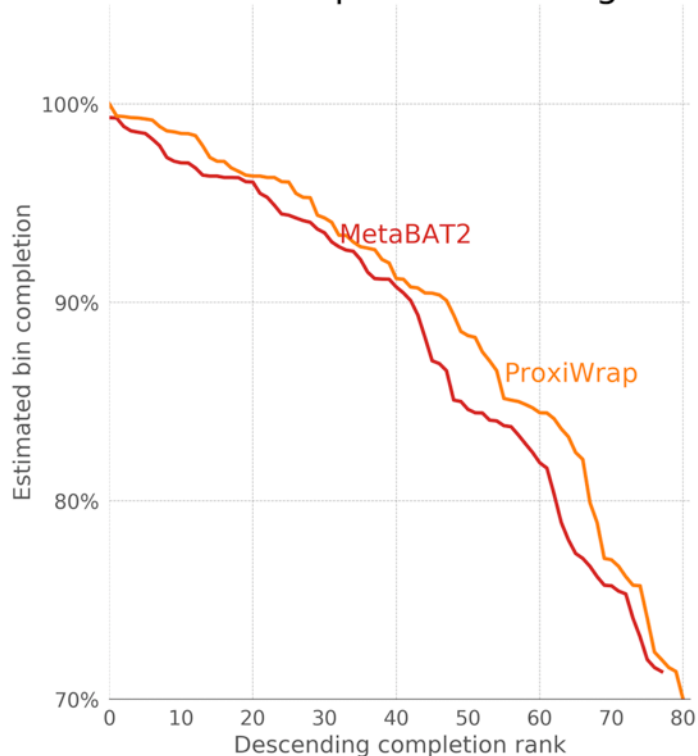
Sequences added and removed by ProxiWrap

Improving intra-cluster connectivity

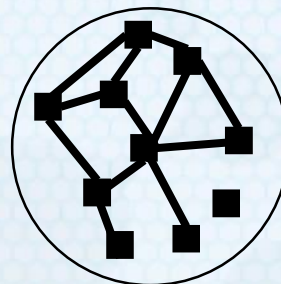
# ProxiWrap™ significantly improves MAG

## completion and provides estimates of MAG validity

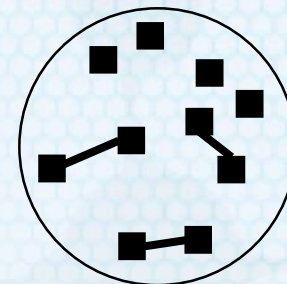
Bin completion ranking



MAG	Completion	Contamination	Connectivity
bin.73	100%	4.2%	88.3%
bin.7	95.8%	2.7%	68.8%
bin.35	98.6%	0.3%	3.2%
bin.111	71.8%	5.7%	0.01%



High-confidence MAG  
(high inter-contig Hi-C  
connectivity)




Low-confidence MAG  
(low inter-contig Hi-C  
connectivity)



# Proximity-Guided Metagenome Assembly™

## Genomes, Strains, Mobile Elements

- No culturing
- No binning/de-replication
- No *a priori* information
- No HMW-DNA
- No special machinery




**PHASE**  
 GENOMICS

Product Announcement


## The ProxiMeta™ Platform

### Kit & Analysis Bundle

ProxiMeta Metagenome Deconvolution Analysis included with kit purchase for limited time.

**New: ProxiMeta 8-Pack Kit**

- Strain-resolution, High-quality genomes
- Associate plasmids, phage, and ARGs with hosts
- Works directly on microbiome samples



For Research Purposes Only

© Phase Genomics 2019

# New 8-pack kits, **ANALYSIS INCLUDED**

# Acknowledgements



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Allergy and  
Infectious Diseases

